

GENES FROM PROPIONIBACTERIUM FREUDENREICHII ENCODING ENZYMES INVOLVED IN VITAMIN B12 BIOSYNTHESIS

Field of Invention

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This invention relates to genes and the proteins (such as enzymes) involved in the biosynthetic pathway of vitamin B₁₂. In particular, the invention provides four novel genes (and their corresponding encoded enzymes), all derived from *Propionibacteria*, in particular *Propionibacterium freudenreichii*. These enzymes are either synthases or transferases, and can be used in the manufacture of vitamin B₁₂.

Introduction

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Vitamin B₁₂ is an important vitamin for humans and animals. It is an essential vitamin, and is obtained from foodstuffs in the human and animal diet. Vitamin B₁₂ is naturally found in animal foods including fish, milk and milk products, eggs, meat and poultry. Certain foodstuffs, for example breakfast cereals, are fortified in vitamin B₁₂, and provide a particularly valuable source of the vitamin for vegetarians.

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Vitamin B₁₂ is used to treat pernicious anaemia and peripheral neuritis, and is also used as a supplement to animal feeds.

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The term vitamin B₁₂ is used to describe compounds of the cobalt corrinoid family, in particular those of the cobalamin group. The most referred to compound of this group is cyanocobalamin and as such the term vitamin B₁₂ is sometimes used to refer to cyanocobalamin. In this specification the term vitamin B₁₂ should be attributed its broad meaning so as to include all the cobalt corrinoids of the cobalamin group, which include in particular cyanocobalamin, hydroxocobalamin, methylcobalamin and 5'-desoxyadenosylcobalamin characterised by a cyano, hydroxyl, methyl or 5'-desoxyadenosyl radical respectively.

Vitamin B₁₂ is produced industrially by microbial fermentation, especially using *Pseudomonas denitrificans*. However, current production levels of vitamin B₁₂ do not always enable cost-effective production of vitamin B₁₂. To increase vitamin B₁₂ productivity, efforts need to be made to improve the fermentation process.

5 The biosynthetic pathway of vitamin B₁₂ in *Pseudomonas denitrificans* has been well characterized²⁴. This has elucidated most of the pathway²⁵. A total of 22 enzymes were purified, and 22 *cob* genes have been identified. The role of some of these genes however is still unknown. It is thought that a closely related, but somewhat different pathway, operates in *Propionibacterium shermanii*.

10 In addition, workers have studied the cobalamine by synthetic pathway in *Salmonella typhimurium*. The *S. typhimurium cob* operon was isolated and cloned into *E. coli*, and that approach gave the new host the ability to make cobalamins *de novo*, an ability which did not previously exist. In terms of patent publications, Blanche of Rhone Poulenc Rorer is referred to concerning biosynthesis methods
15 enabling the preparation of cobalamins.

A total of 14 genes, encoding enzymes responsible for 17 steps of the anaerobic B₁₂ pathway in *Propionibacterium freudenreichii*, have been proposed⁴¹. However, this document does not give any sequences, and expresses only two gene products, allegedly causing methylation. They are expressed in *E. coli*, and there is no
20 disclosed use of these genes to actually produce vitamin B₁₂.

While vitamin B₁₂ has been produced industrially using *Propionibacterium* species, the yields and production levels are not entirely satisfactory, and there is room for improvement. Therefore, research has been undertaken to elucidate the biosynthetic pathway in *Propionibacterium freudenreichii*, and as a result four
25 different genes and enzymes, of the present invention, have been identified. This allows improvements in vitamin B₁₂ yield on an industrial scale.

Summary of the Invention

30 Novel enzymes are now provided which are involved in the biosynthesis of vitamin B₁₂.

At its broadest, the invention in a first aspect relates to a synthase or transferase from a Gram positive bacteria from the Order Actinomycetales, for example family *Propionibacteriaceae*, such as of the genus *Propionibacterium*, such as the species *Propionibacterium freudenreichii*. These enzymes are (e.g. amide) synthases or (e.g. phospho or nucleotidyl) transferases. Preferably they have the activity EC 6.3.1.-, 2.7.7.-, 2.7.8.- or 2.5.1.17.

More specifically, the present invention provides, in a first aspect, an (isolated and/or purified) synthase or transferase polypeptide comprising:

- (i) the amino acid sequence of SEQ ID No: 2, 4, 6 or 8; or
- 10 (ii) a variant of (i) which is a synthase or transferase; or
- (iii) a fragment of (i) or (ii) which is a synthase or transferase.

According to a second aspect of the invention there is provided a polynucleotide which comprises:

- 15 (a) the nucleic acid sequence of SEQ ID No. 1, 3, 5 or 7, or a sequence encoding a polypeptide of the invention;
- (b) a sequence which is complementary to, or which hybridises to, any sequence as defined in (a);
- (c) a fragment of any sequence in (a) or (b);
- (d) a sequence having at least 60, 65 or 70% identity to any sequence as
20 defined in (a), (b) or (c); or
- (e) a sequence that is degenerate as a result of the genetic code to any of the sequences as defined in (a) to (d).

The invention also provides:

- 25 - an (e.g. expression) vector (third aspect) which comprises a polynucleotide of the invention and which may be capable of expressing a polypeptide of the invention;
- a host (fourth aspect), such as cell line or strain comprising a vector of the invention;
- 30 - a method of producing a polypeptide of the invention which method comprises maintaining a cell line or strain of the invention under conditions suitable for obtaining expression of the polypeptide and, if

necessary, isolating the polypeptide; and
- a method of producing vitamin B₁₂ or a precursor thereof (fifth aspect), the method comprising contacting a substrate with a polypeptide or host cell of the invention.

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Brief Description of the Sequences

- SEQ ID No. 1 is a DNA sequence of a first enzyme, an amide synthase, of the invention from *Propionibacterium freudenreichii*;
- 10 SEQ ID No. 2 is the amino acid sequence of the first enzyme (A);
SEQ ID No. 3 is a DNA sequence of a second enzyme, a (phospho and/or nucleotidyl) transferase from the same organism;
SEQ ID No. 4 is the amino acid sequence of the second enzyme (B);
SEQ ID No. 5 is a DNA sequence of a third enzyme, a transferase also from the same
15 organism;
SEQ ID No. 6 is the amino acid sequence of the third enzyme (C);
SEQ ID No. 7 is a DNA sequence of a fourth enzyme, a (nucleotidyl) transferase also from the same organism;
SEQ ID No. 8 is the amino acid sequence of the fourth enzyme (D); and
20 SEQ ID Nos. 9 to 17 are primers.

Detailed Description of the Invention

A. Polynucleotides

- 25 The present invention provides a (e.g. isolated and/or purified) polynucleotide encoding polypeptides of the invention. The present invention thus provides a polynucleotide, preferably encoding a synthase or transferase whose amino acid sequence is set out in SEQ ID No. 2, 4, 6 and/or 8. The present invention further provides a polynucleotide encoding a polypeptide having substantial amino acid
30 sequence homology to the amino acid sequence set out in SEQ ID No. 2, 4, 6 and/or 8. Also included is a polynucleotide selected from:

- (a) a polynucleotide comprising the nucleotide sequence set out in SEQ ID No. 1, 3, 5 and/or 7, or the complement thereof;
- (b) a polynucleotide comprising a nucleotide sequence capable of (e.g. selectively) hybridising to a nucleotide sequence set out in SEQ ID No. 1, 3, 5 or 7, or a fragment thereof;
- (c) a polynucleotide comprising a nucleotide sequence capable of (e.g. selectively) hybridising to the complement of the nucleotide sequence set out in SEQ ID No. 1, 3, 5 or 7, or a fragment thereof; and/or
- (d) a polynucleotide comprising a polynucleotide sequence that is degenerate as a result of the genetic code to a polynucleotide defined in (a), (b) or (c).

Polynucleotides may comprise 2, 3 or more sequences of the invention; for example SEQ ID

Nos. 3 and 5 or 1, 3 and 5 (or variants thereof as defined in any of (b), (c) and (d) above).

A polynucleotide of the invention may also include a polynucleotide which:

- (a) encodes a polypeptide having synthase or transferase activity, which polynucleotide is:
- (1) the coding sequence of SEQ ID No. 1, 3, 5 or 7;
 - (2) a sequence which hybridises selectively to the complement of sequence defined in (1); or
 - (3) a sequence that is degenerate as a result of the genetic code with respect to a sequence defined in (1) or (2); or
- (b) is a sequence complementary to a polynucleotide defined in (a).

Hybridisable sequences

The term "capable of hybridizing" means that the target polynucleotide of the invention can hybridize to a nucleic acid used as a probe (for example the nucleotide sequence set out in SEQ. ID No. 1, 3, 5 or 7, or a fragment thereof or the complement thereof) at a level significantly above background. The invention also includes nucleotide sequences that encode for synthase or transferase or variants thereof as

well as nucleotide sequences that are complementary thereto. The nucleotide sequence may be RNA or DNA and thus includes genomic DNA, synthetic DNA or cDNA). Preferably the nucleotide sequence is a DNA sequence if appropriate and, a cDNA sequence. Typically a polynucleotide of the invention comprises a contiguous sequence of nucleotides which is capable of hybridizing under selective conditions to the coding sequence or the complement of the coding sequence of SEQ ID No. 1, 3, 5 or 7, as appropriate. Such nucleotides can be synthesized according to methods well known in the art¹.

A polynucleotide of the invention can hybridize to the coding sequence or the complement of the coding sequence of SEQ ID No. 1, 3, 5 or 7 (as appropriate) at a level significantly above background. Background hybridization may occur, for example, because of other cDNAs present in a cDNA library. The signal level (e.g. generated by the interaction between a polynucleotide of the invention and the coding sequence or complement of the coding sequence of SEQ ID No. 1, 3, 5 or 7) is typically at least 10 fold, preferably at least 100 fold, or as intense as interactions between other polynucleotides and the coding sequence of SEQ ID No. 1, 3, 5 or 7. The intensity of interaction may be measured, for example, by radiolabelling the probe, e.g. with ³²P. Selective hybridization may typically be achieved using conditions of low stringency (0.3M sodium chloride and 0.03M sodium citrate at about 40°C), medium stringency (for example, 0.3M sodium chloride and 0.03M sodium citrate at about 50°C) or high stringency (for example, 0.3M sodium chloride and 0.03M sodium citrate at about 60°C). Hybridization may be carried out under any suitable conditions known in the art¹ and, as a guide, low stringency can be 2 x SSC at 55°C, medium stringency can be 0.5 to 1.0 x SSC at 60°C and high stringency can be 0.1 or 0.2 x SSC at 60°C or higher (e.g. at 68°C), all at 0.5% SDS.

Modifications

Polynucleotides of the invention may comprise DNA or RNA. They may be single or double stranded. They may also be polynucleotides which include within them one or more synthetic or modified nucleotides. A number of different types of

modifications to polynucleotides are known in the art. These include methylphosphonate and phosphorothioate backbones and/or addition of acridine or polylysine chains at the 3' and/or 5' ends of the molecule. For the purposes of the present invention, it is to be understood that the polynucleotides described herein
5 may be modified by any method available in the art.

It is to be understood that skilled persons may, using routine techniques, make nucleotide substitutions that do not affect the polypeptide sequence encoded by the polynucleotides of the invention to reflect the codon usage of any particular host organism, for example in which the polypeptides of the invention are to be expressed.
10 The coding sequence of SEQ ID No. 1, 3, 5 or 7 may be modified by nucleotide substitutions, for example from or up to 1, 2 or 3 to 10, 25, 50 or 100 substitutions. The polynucleotide may alternatively or additionally be modified by one or more insertions and/or deletions and/or by an extension at either or both ends. The modified polynucleotide generally encodes a polypeptide which has synthase or
15 transferase activity. Degenerate substitutions may be made and/or substitutions may be made which would result in a conservative amino acid substitution when the modified sequence is translated, for example as discussed with reference to polypeptides later.

20 *Homologues.*

A nucleotide sequence which is capable of selectively hybridizing to (e.g. the complement of) the DNA coding sequence of SEQ ID No. 1, 3, 5 or 7 may have at least 50% or 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 98%
25 or at least 99% sequence identity (or homology) to the coding sequence of SEQ ID No. 1, 3, 5 or 7. This may be over a region of at least 20, preferably at least 30, for instance at least 40 or 50 such as at least 60 or 80, more preferably at least 100, 200, 400, 500 or even 600 contiguous nucleotides or optimally over the full length of SEQ ID No. 1, 3, 5 or 7. For individual sequences the sequence identity may be:

- 30 (a) for SEQ ID No. 1, at least 85% or 90%;
(b) for SEQ ID No. 3, at least 70%;

(c) for SEQ ID No. 5, at least 90% or 95%; and/or

(d) for SEQ ID No. 7, at least 90%, 95% or 98%.

Any combination of the above mentioned degrees of homology and minimum sized may be used to define polynucleotides of the invention, with the more stringent combinations (i.e. higher homology over longer lengths) being preferred. Thus for example a polynucleotide which is at least 80% or 90% homologous over 25, preferably over 30 nucleotides forms one aspect of the invention, as does a polynucleotide which is at least 90% homologous over 40 nucleotides.

Homologues of polynucleotide (or protein) sequences typically have at least 70% homology, preferably at least 80, 90%, 95%, 97% or 99% homology, for example over a region of at least 15, 20, 30, 100 more contiguous nucleotides (or amino acids). The homology may be calculated on the basis of amino acid identity (sometimes referred to as "hard homology"). Identity or homology is usually calculated on the basis of the entire sequence unless otherwise specified.

For example the UWGCG Package provides the BESTFIT program which can be used to calculate homology (for example used on its default settings⁵). The PILEUP and BLAST algorithms can be used to calculate homology or line up sequences (such as identifying equivalent or corresponding sequences, for example on their default settings^{6,7}).

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pair (HSPs) by identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold^{6,7}. These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Extensions for the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring

residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLAST program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix⁸ alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of
5 both strands.

The BLAST algorithm performs a statistical analysis of the similarity between two sequences⁹. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by
10 chance. For example, a sequence is considered similar to another sequence if the smallest sum probability in comparison of the first sequence to the second sequence is less than about 1, preferably less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

Fragments, homologues and other variants may be at least 500 or 550
15 nucleotides in length (e.g. for SEQ. ID. No. 7) and may encode at least (e.g. the first) 170, 180 or 200 amino acids of the corresponding protein.

Primers and Probes

20 Polynucleotides of the invention include and may be used as a primer, e.g. a PCR primer, a primer for an alternative amplification reaction, a probe, or the polynucleotides may be cloned into vectors. Such primers, probes and other fragments will be at least or up to 20, for example at least 25, 30 or 40 nucleotides in length. They will typically be up to 40, 50, 60, 70, 100, 150, 200 or 300 nucleotides
25 in length, or even up to the same number or a few nucleotides (such as 5 or 10 nucleotides) short of the coding sequence of SEQ ID No. 1, 3, 5 or 7.

In general, primers will be produced by synthetic means, involving a step-wise manufacture of the desired nucleic acid sequence one nucleotide at a time. Techniques for accomplishing this using automated techniques are readily available
30 in the art.

Longer polynucleotides will generally be produced using recombinant means,

for example using PCR (polymerase chain reaction) cloning techniques. This will involve making a pair of primers (e.g. of about 15-30 nucleotides) to a region of the synthase or transferase which it is desired to clone, bringing the primers into contact with mRNA or cDNA obtained from a target (e.g. yeast, bacterial, plant, prokaryotic or fungal) cell, preferably of an a bacterial, e.g. *Propionibacterium* strain, performing a polymerase chain reaction under conditions which bring about amplification of the desired region, isolating the amplified fragment (e.g. by purifying the reaction mixture on an agarose gel) and recovering the amplified DNA. The primers may be designed to contain suitable restriction enzyme recognition sites so that the amplified DNA can be cloned into a suitable cloning vector.

Such techniques may be used to obtain all or part of the synthase or transferase sequence described herein. Genomic clones corresponding to the cDNA of SEQ ID No. 1, 3, 5 or 7 or the synthase or transferase gene containing, for example, introns and promoter regions are within the invention also and may also be obtained in an analogous manner (e.g. recombinant means, PCR, cloning techniques), starting with genomic DNA from a fungal, yeast, bacterial plant or prokaryotic cell.

The polynucleotides or primers may carry a revealing label, e.g. a radioactive or non-radioactive label. Suitable labels include radioisotopes such as ^{32}P or ^{35}S , enzyme labels, or other protein labels such as biotin. Such labels may be added to polynucleotides or primers of the invention and may be detected using techniques known *per se*.

Polynucleotides, labelled or unlabelled may be used in nucleic acid-based tests for detecting or sequencing a synthase or transferase or a variant thereof in a (e.g. bacterial) sample. Such tests for detecting generally comprise bringing a (e.g. bacterial) sample (suspected of) containing DNA into contact with a probe or primer of the invention under hybridizing conditions and detecting any duplex formed between the probe and nucleic acid in the sample. Such detection may be achieved using techniques such as PCR or by immobilizing the probe on a solid support, removing nucleic acid in the sample which is not hybridized to the probe, and then detecting nucleic acid which was hybridized to the probe. Alternatively, the sample nucleic acid may be immobilized on a solid support, and the amount of probe bound

to such a support can be detected.

The probes of the invention may conveniently be packaged in the form of a test kit in a suitable container. In such kits the probe may be bound to a solid support where the assay format for which the kit is designed requires such binding. The kit may also contain suitable reagents for treating the sample to be probed, hybridizing the probe to nucleic acid in the sample, control reagents, instructions, and the like.

Preferably, the polynucleotide of the invention is obtainable from the same organism as the polypeptide, such as a bacteria, in particular a bacteria of the family *Mycobacteriaceae*, preferably of the genus *Propionibacterium*.

The polynucleotides of the invention also include variants of the sequence of SEQ ID No. 1, 3, 5 or 7 which have synthase or transferase activity. Variants may be formed by additions, substitutions and/or deletions and may have the ability to act as a synthase or transferase, or have the activity EC 6.3.1-, 2.7.7 (or 8).- or 2.5.1.17.

15 *Production of polynucleotides*

Polynucleotides which do not have 100% identity with SEQ ID No. 1, 3, 5 or 7 but fall within the scope of the invention can be obtained in a number of ways. Thus variants of the sequences described herein may be obtained for example by probing genomic DNA libraries made from a range of organisms, for example those discussed as sources of the polypeptides of the invention. In addition, other bacterial or prokaryotic homologues may be obtained and such homologues and fragments thereof in general will be capable of hybridising to SEQ ID No. 1, 3, 5 or 7. Such sequences may be obtained by probing cDNA libraries or genomic DNA libraries from other species, and probing such libraries with probes comprising all or part of SEQ ID. 1, 3, 5 or 7 under conditions of medium to high stringency (as described earlier). Nucleic acid probes comprising all or part of SEQ ID No. 1, 3, 5 or 7 may be used to probe cDNA libraries from other species, such as those described as sources for the polypeptides of the invention.

Species homologues may also be obtained using degenerate PCR which will use primers designed to target sequences within the variants and homologues encoding

conserved amino acid sequences. The primers can contain one or more degenerate positions and will be used at stringency conditions lower than those used for cloning sequences with single sequence primers against known sequences.

Alternatively, such polynucleotides may be obtained by site directed
5 mutagenesis of the
synthase or transferase sequences or variants thereof. This may be useful where for example silent codon changes are required to sequences to optimise codon preferences for a particular host cell in which the polynucleotide sequences are being expressed. Other sequence changes may be desired in order to introduce restriction
10 enzyme recognition sites, or to alter the property or function of the polypeptides encoded by the polynucleotides.

The invention includes double stranded polynucleotides comprising a polynucleotide of the invention and its complement.

The present invention also provides polynucleotides encoding the polypeptides
15 of the invention described below. Since such polynucleotides will be useful as sequences for recombinant production of polypeptides of the invention, it is not necessary for them to be capable of hybridising to the sequence of SEQ ID No. 1, 3, 5 or 7, although this will generally be desirable. Otherwise, such polynucleotides may be labelled, used, and made as described above if desired.

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B. Polypeptides

The present invention relates to an (e.g. (substantially) purified and/or isolated) synthase or transferase or variant thereof as defined later. The polypeptides of the
25 invention may consist essentially of the amino acid sequence of SEQ ID No. 2, 4, 6 or 8 or of a variant of that sequence. Polypeptides may also be encoded by a polynucleotide of the invention as described above.

A polypeptide of the invention may be in an isolated or a substantially purified form. It will be understood that the polypeptide may be mixed with carriers or
30 diluents which will not interfere with the intended purpose and/of function of the polypeptide and still be regarded as substantially isolated. It will generally comprise

the polypeptide in a preparation in which more than 20%, e.g. more than 30%, 40%, 50%, 80%, 90%, 95% or 99%, by weight of the polypeptide in the preparation is a polypeptide of the invention. Routine methods can be employed to purify and/or synthesise the proteins according to the invention¹. For some formulations (e.g. for non-pharmaceutical uses) the amount of polypeptide present may be small, for example from 0.01 to 10%, such as from 0.1 to 5%, or 2% or even from 0.2 to 1%.

Preferably, the polypeptide of the invention is obtainable from a microorganism, for example one which possesses a gene encoding an enzyme with synthase or transferase activity. More preferably the microorganism is bacterial, such as Gram positive bacteria. The microorganism may be of the Phylum or class Actinobacteria, e.g. Subclass *Adinobacteridae*. Preferably the microorganism is of Order *Actinomycetales*, such as Sub order *Propionibacterineae* and optimally from the family *Propionibacteriaceae*. Preferred organisms are thus of the genus *Propionibacterium*, such as of the species *Propionibacterium freudenreichii*. Preferably the microorganism is capable of producing or synthesising vitamin B12.

Activity

A polypeptide of the invention can have one or more of the following features, namely it:

- (1) possesses synthase or transferase activity;
- (2) acts as an amide synthase or a phospho-, nucleotidyl- or aryl-transferase;
- (3) catalyses at least one step in the vitamin B₁₂ biosynthetic pathway;
- (4) has an activity within EC 6.3.1-, EC 2.7.7-, EC 2.7.8- or EC 2.5.1.17;
- (5) has a length of 150 or 170 to 270 or 300 amino acids or from 800 or 840 to 880 or 920 amino acids;
- (6) is a cobyrinic acid -a,c-diamide synthase, a cobinamide kinase, a cobinamide phosphate guanyl transferase, a cobalamin (5'-phosphate) synthase and/or an adenosyl transferase;
- (7) acts on a substrate, or produces a product, which comprises:
 - (i) a corrin core or ring system;

(ii) up to 4 aryl, optionally pyrrole, rings;

(iii) a tetrapyrrole ring system and/or a transition metal (e.g. cobalt) atom;

and/or

(iv) an amide, phosphate, guanidyl, aryl or adenosyl moiety or group;

5 and/or

(8) catalyses amidation, phosphorylation, nucleotidylation, arylation, ribazole or adenosyl addition and/or adenosylation.

A table of the main characteristics of the polypeptides of the invention is given below.

10

Designation / reaction	DNA seq No. (length, nucs)	Protein seq No.	Seq length (amino acids)	Enzyme activity/ activities	Substrate	Product (formula)	Enzyme class/type	Gene refere No. (name)
A	1 (2586)	2	861	cobyrrinic acid a,c diamide synthase (EC 6.6.3.-)	Cobyrrinic acid (I)	Cobyrrinic a,c diamide (IB) (with intermediate coobyrrinic acid c-amide, IA)	(amide) synthase	PFR 1 (cobA)
B (B1, B2)	3 (657)	4	218	cobinamide kinase (EC 2.7.1.-)	Adenosyl cobinamide (II)	Adenosinyl cobinamide phosphate (IIA)	(phospho) transferase	PFR 1 (cobU)
				cobinamide phosphate guanyl transferase (EC 2.7.7.-)	Adenosyl cobinamide phosphate (IIA)	Adenosyl-GDP-cobamide (IIB)	(nucleotidyl) transferase	
C	5 (780)	6	256	cobalamin (5'-phosphate) synthase (EC 2.7.8.-)	Adenosyl-GDP-cobamide (IIB)	Adenosyl-5,6-dimethyl benzimidazolyl cobamide, (adenosyl cobalamin i.e. vitamin B ₁₂ (IIC))	(aryl) transferase	PFR 1 (cobS)
D	7 (603)	8	200	(cob(1)alamin) adenosyl transferase (EC 2.5.1.17)	Cobyrrinic acid a, c diamide (IB)	Adenosyl coobyrrinic acid a, c diamide (IC)	(nucleotidyl) transferase	PFR 1

Variants and Homologues

15

A polypeptide of the invention can comprise the amino acid sequence set out in

SEQ ID No. 2, 4, 6 or 8 (or a variant thereof, such as) a substantially homologous sequence or a fragment of either sequence and can have synthase or transferase activity. In general, the naturally occurring amino acid sequence shown in SEQ ID No. 2, 4, 6 or 8 is preferred.

- 5 In particular, the polypeptide of the invention may comprise:
- (a) the polypeptide sequence of SEQ ID No. 2, 4, 6 or 8;
 - (b) a naturally occurring variant or species homolog, paralog or ortholog thereof; or
 - (c) a protein with at least 70, at least 80, at least 90, at least 95, at least 98 or at
10 least 99% sequence identity to (a) or (b).

A variant may be one that occurs naturally, for example in fungal, bacteria, yeast or plant cells and which can function in a substantially similar manner to the protein of SEQ ID No. 2, 4, 6 or 8, for example it has synthase or transferase activity. Similarly a species homolog of the protein will be the equivalent protein which
15 occurs naturally in another species and which can function as a synthase or transferase enzyme. Variants include allelic variants either from the same strain as the polypeptide of the invention or from a different strain, but of the same genus, or of the same species.

Variants and species homologues can be obtained by following the procedures
20 described herein for the production of the polypeptide of SEQ ID No. 2, 4, 6 or 8 and performing such procedures on a suitable cell source, for example a bacterial, yeast, fungal or plant cell. It will also be possible to use a probe as defined above to probe libraries made from yeast, bacterial, fungal or plant cells in order to obtain clones including the variants or species homology. The clones can be manipulated by
25 conventional techniques to generate a polypeptide of the invention which can then be produced by recombinant or synthetic techniques known *per se*.

The polypeptide of the invention preferably has at least 70% sequence identity to any of the proteins of SEQ ID No. 2, 4, 6 or 8, more preferably at least 80%, at least 90%, at least 95%, at east 97% or at least 99% sequence identity thereto, for example
30 for each sequence over a region of at least 60, at least 100, 150, 200, 250 or 300 (or even 500, 600, 700 or 800) contiguous amino acids or over the full length of SEQ ID

No. 2, 4, 6 or 8. For individual sequences the sequence identity may be:

- (a) for SEQ ID No. 2, at least 55%, 60% or 65%;
- (b) for SEQ ID No. 4, at least 50%, 55% or 60%;
- (c) for SEQ ID No. 6, at least 40%, 45% or 50%; and/or
- 5 (d) for SEQ ID No. 8, at least 90%, 95%, 98% or 99% (e.g. over, or the sequence being longer than, at least 150, 170, 200 or 230 amino acids).

The sequence of the polypeptide of SEQ ID No. 2, 4, 6 or 8 and of variants and species homologs can thus be modified to provide polypeptides of the invention.

10 Amino acid substitutions may be made, for example from or up to 1, 2 or 3 to 10, 20, 30, 50 or 100 substitutions. The same number of deletions and insertions may also be made. These changes may be made outside regions critical to the function of the polypeptide and so may still result in an active enzyme. The modified polypeptide generally retains activity as a synthase or transferase.

15 Polypeptides of the invention include fragments of the above mentioned full length polypeptides and of variants thereof, including fragments of the sequence set out in SEQ ID No. 2, 4, 6 or 8. Such fragments typically retain activity as a synthase or transferase. Fragments may be at least 50, 100, 150, 200 or 250 amino acids long or may be this number of amino acids short of the full length sequence (as shown in
20 SEQ ID No. 2, 4, 6 or 8).

Polypeptides of the invention can if necessary be produced by synthetic means although usually they will be made recombinantly as described below. They may be modified for example by the addition of histidine residues or a T7 tag to assist their identification or purification or by the addition of a signal sequence to promote their
25 secretion from a cell.

The term "variants" refers to polypeptides which have the same essential character or basic biological functionality as the synthase or transferase, and include allelic variants. The essential character of synthase is that it is an enzyme that exhibits EC 6.3.-.-. (e.g. EC 6.3.1.-.) activity or can add an amine group to a substrate
30 (e.g. amidate). For a transferase, this is an enzyme that exhibits EC 2.7.-.-. (e.g. EC 2.7.7 or 8.-) or EC 2.5.-.-. (e.g. EC 2.5.1.-, such as EC 2.5.1.17) activity and can

transfer a substituent or chemical moiety from one compound to another. Preferably a variant polypeptide has the same activity. A polypeptide having the same essential character as may be identified by performing a substrate degradation assay.

Variants of SEQ ID No. 2, 4, 6 or 8 also include sequences which vary from
 5 SEQ ID No. 2, 4, 6 or 8 but which are not necessarily derived from the naturally occurring enzyme. These variants may be described as having a percentage homology to SEQ ID No. 2, 4, 6 or 8 or having a number of substitutions within this sequence. Alternatively a variant may be encoded by a polynucleotide which hybridizes to SEQ ID No. 1, 3, 5 or 7.

10 The variants can be defined in a similar manner to the variants of SEQ ID No. 1, 3, 5 or 7. Thus the variants may comprise variant sequences derived from other bacterial strains, e.g. *Propionibacterium*. Other variants can be identified from other strains by looking for synthase or transferase activity and cloning and sequencing as before. Variants may include the deletion, modification or addition of single amino
 15 acids or groups of amino acids within the protein sequence, as long as the peptide maintains the basic biological functionality. e.g. a synthase or transferase.

Conservative substitutions may be made, for example according to the following Table. Amino acids in the same block in the second column and preferably in the
 20 same line in the third column may be substituted for each other. Preferably substitutions do not affect the folding or activity of the polypeptide.

ALIPHATIC	Non-polar	G A P
		I L V
	Polar-uncharged	C S T M
		N Q
	Polar-charged	D E
		K R
AROMATIC		H F W Y

Shorter polypeptide sequences are within the scope of the invention. For
 example, a peptide of at least 50 amino acids or up to 60, 70, 80, 100, 150, 200, 400,
 500, 600 or 700 amino acids in length is considered to fall within the scope of the
 25 invention as long as it demonstrates the basic biological functionality of the synthase

or transferase. In particular, but not exclusively, this aspect of the invention encompasses the situation when the protein is a fragment of the complete protein sequence and may comprise or represent a substrate binding region, cleaving and/or transferring region.

5

Modifications

Polypeptides of the invention may be chemically modified, e.g. post-translationally

10 modified. For example, they may comprise modified amino acid residues. They may also be modified by the addition of histidine residues (to assist their purification) or by the addition of a signal sequence (to promote insertion into the cell membrane). The polypeptide may have one or more (N) amino- or (C) carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of
15 up to about 20-25 residues, or a (small) extension that facilitates purification, such as a poly-histidine or T7 tag, an antigenic epitope or a (e.g. maltose) binding domain¹⁴ (e.g. at the C-terminus). These extensions may or may not be added via a linker.

A polypeptide of the invention may be labelled with a revealing label. The revealing label may be any suitable label which allows the polypeptide to be detected.

20 Suitable labels include radioisotopes, e.g. ¹²⁵I, ³⁵S, enzymes, antibodies, polynucleotides and linkers such as biotin.

The polypeptide may be modified to include non-naturally occurring amino acids or to increase the stability of the polypeptide. When the peptide is produced by synthetic means, such amino acids may be introduced during production. The
25 peptide may also be modified following either synthetic or recombinant production.

The polypeptides of the invention may also be produced using, or comprise, (one or more) D-amino acids.

A number of side chain modifications are known in the art and may be made to the side chains of the proteins or peptides of the present invention. Such
30 modifications include, for example, modifications of amino acids by reductive alkylation by reaction with an aldehyde followed by reduction with NaBH₄,

amidination with methylacetimidate or acylation with acetic anhydride.

The sequences provided by the present invention may also be used as starting materials for the construction of "second generation" enzymes. "Second generation" enzymes are ones altered by mutagenesis techniques (e.g. site-directed mutagenesis), which have properties that differ from those of the wild-type or recombinant enzymes such as those produced by the present invention. For example, the temperature or pH optimum, specific activity, substrate affinity or thermostability may be altered so as to be better suited for application in a defined process.

Amino acids essential to activity, and therefore preferably subject to substitution, may be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis¹⁰. In the latter technique mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity (e.g. synthase or transferase activity) to identify amino acid residues that are critical to the activity of the molecule. Sites of enzyme-substrate interaction can also be determined by analysis of crystal structure as determined by such techniques as nuclear magnetic resonance, crystallography or photo-affinity labelling^{11,12,13} or molecular modelling.

The use of yeast and fungal host cells may provide any post-translational modifications (e.g. proteolytic processing, myristilation, glycosylation, truncation, and tyrosine, serine or threonine phosphorylation) influencing biological activity on recombinant expression products of the invention.

Polypeptides of the invention may be provided in a form such that they are outside their natural cellular environment. Thus, they may be substantially isolated or purified, as discussed above, or in a cell in which they do not occur in nature, e.g. a cell of other bacterial species, animals, yeast or fungi.

C. Recombinant Aspects.

The invention also provides vectors comprising a polynucleotide of the invention, including cloning and expression vectors, and methods of growing, transforming or transfecting such vectors in a suitable host cell, for example under

conditions in which expression of a polypeptide of the invention occurs. Provided also are host cells comprising a polynucleotide or vector of the invention wherein the polynucleotide is heterologous to the genome of the host cell. The term "heterologous", usually with respect to the host cell, means that the polynucleotide does not naturally occur in the genome of the host cell or that the polypeptide is not naturally produced by that cell. Preferably, the host cell is a bacterial cell, for example a (e.g. Gram positive) cell of the family *Propionibacteriaceae*.

Polynucleotides of the invention can be incorporated into a recombinant replicable vector, for example a cloning or expression vector. The vector may be used to replicate the nucleic acid in a compatible host cell. Thus in a further embodiment, the invention provides a method of making polynucleotides of the invention by introducing a polynucleotide of the invention into a replicable vector, introducing the vector into a compatible host cell, and growing the host cell under conditions which bring about replication of the vector. The vector may be recovered from the host cell. Suitable host cells are described below in connection with expression vectors.

Vectors

The polynucleotide of the invention may be inserted into an expression cassette. The vector into which the expression cassette or polynucleotide of the invention is inserted may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of the vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e. a vector which exists as an extra-chromosomal entity, the replication of which is independent of chromosomal replication, e.g. a plasmid. Alternatively, the vector may be one which, when introduced into a host cell, is integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

Preferably, a polynucleotide of the invention in a vector is operably linked to a regulatory sequence which is capable of providing for the expression of the coding

sequence by the host cell, i.e. the vector is an expression vector. The term "operably linked" refers to a juxtaposition wherein the components described are in a relationship permitting them to function in their intended manner. A regulatory sequence such as a promoter, enhancer or other expression regulation signal

5 "operably linked" to a coding sequence is positioned in such a way that expression of the coding sequence is achieved under condition compatible with the control sequences.

The vector may be a plasmid, cosmid, virus or phage vector, usually provided with an origin of replication, optionally a promoter for the expression of the

10 polynucleotide and optionally an enhancer and/or a regulator of the promoter. A terminator sequence may be present, as may be a polyadenylation sequence. The vector may contain one or more selectable marker genes, for example an ampicillin resistance gene (in the case of a bacterial plasmid) or a neomycin resistance gene (for a mammalian vector). Vectors may be used *in vitro*, for example for the production

15 of RNA or used to transfect or transform a host cell. They may comprise two or more polynucleotides of the invention, for example for overexpression.

The vector may comprise two, three or all four polynucleotides of the invention, in other words at least two, three or four polynucleotide sequences that encode the four polypeptides of the invention (SEQ ID Nos. 2, 4, 6 and 8, or variants (fragments

20 or substantially homologous sequences thereof), as defined earlier). Preferred combinations include the sequences encoding protein B (cobU) and protein C (cobS), optionally with the third protein A (cobA/cbiA). Thus, the vector may comprise SEQ ID Nos. 1, 3, 5 and/or 7, or fragments thereof, or sequences that hybridize thereto, as defined earlier in the specification.

25 Preferably at least 2, 3 and (optimally) 4 of the polynucleotides are in the same operon, for example operon C. They may be arranged so that the vector (or eventual host) comprises an operon, or sequence, comprising sequences encoding one or more of the following enzymes, in the order: (nucleotidyl, transferase, (amide) synthase, (phospho) transferase and/or (nucleotidyl) transferase, (aryl) transferase. Thus, if all

30 four polynucleotides are present, then the order in which they are arranged is preferably SEQ ID Nos. 7, 1, 3, 5 (or variants of these sequences as previously

defined).

The DNA sequence encoding the polypeptide is preferably introduced into a suitable host as part of an expression cassette (or construct) in which the DNA sequence is operably linked to expression signals which are capable of directing
5 expression of the DNA sequence in the host cells. For transformation of the suitable host with the expression construct transformation procedures are available which are well known to the skilled person^{3,4}. The expression construct can be used for transformation of the host as part of a vector carrying a selectable marker, or the expression construct may be co-transformed as a separate molecule together with the
10 vector carrying a selectable marker. The vector may comprise one or more selectable marker genes.

Preferred selectable markers^{15,16} include but are not limited to those that complement a defect in the host cell or confer resistance to a drug. They include e.g. versatile marker genes that can be used for transformation of bacteria (e.g. *E. coli*),
15 most filamentous fungi and yeasts such as acetamidase genes or cDNAs (the *amdS*, *niaD*, *facA* genes or cDNAs from *A.nidulans*, *A.oryzae*, or *A.niger*), or genes providing resistance to antibiotics like G418, hygromycin, bleomycin, kanamycin, phleomycin or benomyl resistance (*benA*). Alternatively, specific selection markers can be used such as auxotrophic markers which require corresponding mutant host
20 strains: e.g. *URA3* (from *S.cerevisiae* or analogous genes from other yeasts), *pyrG* or *pyrA* (from *A.nidulans* or *A.niger*), *argB* (from *A.nidulans* or *A.niger*) or *trpC*. In a preferred embodiment the selection marker is deleted from the transformed host cell after introduction of the expression construct so as to obtain transformed host cells capable of producing the polypeptide which are free of selection marker genes^{21,22}.

25 Other markers include ATP synthetase, subunit 9 (*oliC*), orotidine-5'-phosphate-decarboxylase (*pvrA*), the bacterial G418 resistance gene (this may also be used in yeast, but not in fungi), the ampicillin resistance gene (*E. coli*), the neomycin resistance gene (*Bacillus*) and the *E. coli uidA* gene, coding for β -glucuronidase (GUS). Vectors may be used *in vitro*, for example for the production of RNA or used
30 to transfect or transform a host cell.

For most (filamentous) fungi, yeast or bacteria, the vector or expression

construct is preferably integrated, such as in the genome of the host cell in order to obtain stable transformants. However, for certain yeasts also suitable episomal vectors are available into which the expression construct can be incorporated for stable and high level expression, examples thereof include vectors derived from the 2 μ and pKD1 plasmids of *Saccharomyces* and *Kluyveromyces*, respectively, or vectors containing an AMA sequence (e.g. AMA1 from *Aspergillus*^{3,20}). In case the expression constructs are integrated in the host cell's genome, the constructs are either integrated at random loci in the genome, or at predetermined target loci using homologous recombination, in which case the target loci preferably comprise a highly expressed gene. A highly expressed gene is a gene whose mRNA can make up at least 0.01% (w/w) of the total cellular mRNA, e.g. under induced conditions, or alternatively, a gene whose gene product can make up at least 0.2% (w/w) of the total cellular protein (or, in case of a secreted gene product, can be secreted to a level of at least 0.05g/l). A number of examples of suitable highly expressed genes are provided below.

A vector or expression construct for a given host cell may comprise the following elements operably linked to each other in a consecutive order from the 5'-end to 3'-end relative to the coding strand of the sequence encoding the polypeptide of the first invention:

- (1) a promoter sequence capable of directing transcription of the DNA sequence encoding the polypeptide in the given host cell;
- (2) optionally, a signal sequence capable of directing secretion of the polypeptide from the given host cell into a culture medium;
- (3) a DNA sequence encoding a mature and preferably active form of the polypeptide; and preferably also
- (4) a transcription termination region (terminator) capable of terminating transcription downstream of the DNA sequence encoding the polypeptide.

Downstream of the DNA sequence encoding the polypeptide there may be a 3' untranslated region containing one or more transcription termination sites (e.g. a terminator). The origin of the terminator is less critical. The terminator can e.g. be

native to the DNA sequence encoding the polypeptide. However, preferably a yeast terminator is used in yeast host cells, a filamentous fungal terminator is used in filamentous fungal host cells and a bacterial terminator in bacterial cells. More preferably, the terminator is endogenous to the host cell (in which the DNA sequence encoding the polypeptide is to be expressed).

Enhanced expression of the polynucleotide encoding the polypeptide of the invention may also be achieved by the selection of heterologous regulatory regions, e.g. promoter, and/or terminator regions, which may serve to increase expression and, if desired, secretion levels of the protein of interest from the expression host and/or to provide for the inducible control of the expression of the polypeptide of the invention.

Aside from the promoter native to the gene encoding the polypeptide of the invention, other promoters may be used to direct expression of the polypeptide of the invention. The promoter may be selected for its efficiency in directing the expression of the polypeptide of the invention in the desired expression host.

Promoters/enhancers and other expression regulation signals may be selected to be compatible with the host cell for which the expression vector is designed. For example prokaryotic promoters may be used, in particular those suitable for use in *E. coli* strains. When expression is carried out in mammalian cells, mammalian promoters may be used. Tissues-specific promoters, for example hepatocyte cell-specific promoters, may also be used. Viral promoters may also be used, for example the Moloney murine leukaemia virus long terminal repeat (MMLV LTR), promoterous sarcoma virus (RSV) LTR promoter, SV40 (e.g. large T antigen) promoter, human cytomegalovirus (CMV) IE promoter, herpes simplex virus promoters or adenovirus promoters, HSV promoters such as the HSV IE promoters), or HPV promoters, particularly the HPV upstream regulatory region (URR). Yeast promoters include *S. cerevisiae* GAL4 and ADH promoters, the *S. pombe* *nmt 1* and *adh* promoter. Mammalian promoters include the metallothionein promoter which may be induced in response to heavy metals such as cadmium and β -actin promoters. Tissue-specific promoters, in particular endothelial or neuronal cell specific promoters (for example the DDAHI and DDAHII promoters), are especially preferred.

A variety of promoters^{15,16} can be used that are capable of directing transcription in the host cells of the invention. Preferably the promoter sequence is derived from a highly expressed gene as previously defined. Examples of preferred highly expressed genes from which promoters are preferably derived and/or which are comprised in preferred predetermined target loci for integration of expression constructs, include but are not limited to genes encoding glycolytic enzymes such as triose-phosphate isomerases (TPI), glyceraldehyde-phosphate dehydrogenases (GAPDH), phosphoglycerate kinases (PGK), pyruvate kinases (PYK), alcohol dehydrogenases (ADH), as well as genes encoding amylases, glucoamylases, proteases, xylanases, cellobiohydrolases, β -galactosidases, alcohol (methanol) oxidases, elongation factors and ribosomal proteins. Specific examples of suitable highly expressed genes include e.g. the *LAC4* gene from *Kluyveromyces* sp., the methanol oxidase genes (*AOX* and *MOX*) from *Hansenula* and *Pichia*, respectively, the glucoamylase (*glaA*) genes from *A.niger* and *A.awamori*, the *A.oryzae* TAKA-amylase gene, the *A.nidulans gpdA* gene and the *T.reesei* cellobiohydrolase genes.

Examples of strong constitutive and/or inducible promoters which are preferred for use in fungal expression hosts^{15,16} are those which are obtainable from the fungal genes for xylanase (*xlnA*), phytase, ATP-synthetase, subunit 9 (*oliC*), triose phosphate isomerase (*tpi*), alcohol dehydrogenase (*AdhA*), α -amylase (*amy*), amyloglucosidase (AG - from the *glaA* gene), acetamidase (*amdS*) and glyceraldehyde-3-phosphate dehydrogenase (*gpd*) promoters.

Examples of strong yeast promoters are those obtainable from the genes for alcohol dehydrogenase, lactase, 3-phosphoglycerate kinase and triosephosphate isomerase.

Examples of strong bacterial promoters are the α -amylase and *SPo2* promoters as well as promoters from extracellular protease genes.

Promoters suitable for plant cells include napaline synthase (*nos*), octopine synthase (*ocs*), mannopine synthase (*mas*), ribulose small subunit (*rubisco ssu*), histone, rice actin, phaseolin, cauliflower mosaic virus (CMV) 35S and 19S and circovirus promoters. All these promoters are readily available in the art.

The vector may further include sequences flanking the polynucleotide giving rise

to RNA which comprise sequences homologous to eukaryotic genomic sequences, preferably mammalian genomic sequences, or viral genomic sequences. This will allow the introduction of the polynucleotides of the invention into the genome of eukaryotic cells or viruses by homologous recombination. In particular, a plasmid
5 vector comprising the expression cassette flanked by viral sequences can be used to prepare a viral vector suitable for delivering the polynucleotides of the invention to a mammalian cell. Other examples of suitable viral vectors include herpes simplex viral vectors^{18,19} and retroviruses, including lentiviruses, adenoviruses, adeno-associated viruses and HPV viruses (such as HPV-16 or HPV-18). Gene transfer
10 techniques using these viruses are known to those skilled in the art. Retrovirus vectors for example may be used to stably integrate the polynucleotide giving rise to the antisense RNA into the host genome. Replication-defective adenovirus vectors by contrast remain episomal and therefore allow transient expression.

The vector may contain a polynucleotide of the invention oriented in an
15 antisense direction to provide for the production of antisense RNA. This may be used to reduce, if desirable, the levels of expression of the polypeptide.

For bacteria, specialised vectors can be employed, for example an expression vector or a plasmid. Suitable vectors and expression systems for *Propionibacteria* are known in the art^{27,28,30}. For example one can use a plasmid
20 from another *Propionibacterium* species, such as *P. acidipropionici*. This plasmid can be used to prepare a shuttle vector (such as pPK705) containing one or more of the six open reading frames of the *P. acidipropionici* plasmid. The vector may contain a drug marker, such as a hygromycin B resistant gene. This vector has been able to successfully transform *Propionibacterium freudenreichii* subspecies
25 *shermanii*. Transformation may be by electroporation.

Several promoters particularly suitable for *Propionibacterium* can be employed, in particular from *Propionibacterium freudenreichii* subspecies *shermanii*. These include the *Propionibacterium* bacterium promoters P4 and P138.

Alternatively or in addition, one may use one or more endogenous plasmids of
30 *Propionibacterium*, or vectors derived from such a plasmid, in order to express a preferably heterologous protein in bacteria. Such plasmids and vectors are known in

the art²⁹ and may be based on plasmids from *Propionibacterium* bacterium LMG16545 (deposited under accession nos. CBS 101022 and CBS 101023).

Host cells and Expression

5 In a further aspect the invention provides a process for preparing a polypeptide according to the invention which comprises cultivating a host cell (e.g. transformed or transfected with an expression vector as described above) under conditions to provide for expression (by the vector) of a coding sequence encoding the polypeptide, and optionally recovering the expressed polypeptide. Polynucleotides of
10 the invention can be incorporated into a recombinant replicable vector, e.g. an expression vector. The vector may be used to replicate the nucleic acid in a compatible host cell. It may contain at least one copy (such as multiple copies) of the polynucleotide of the invention. Thus in a further embodiment, the invention provides a method of making a polynucleotide of the invention by introducing a
15 polynucleotide of the invention into a replicable vector, introducing the vector into a compatible host cell, and growing the host cell under conditions which bring about the replication of the vector. The vector may be recovered from the host cell. Suitable host cells include bacteria, preferably Gram positive e.g. of the family *Propionibacteriaceae*. Others include *E. coli*, yeast, mammalian cell lines and other
20 eukaryotic cell lines, for example insect cells such as Sf9 cells and (e.g. filamentous) fungal cells.

The polypeptide can be produced as a (secreted) protein in which case the DNA sequence encoding a mature form of the polypeptide in the expression construct is operably linked to a DNA sequence encoding a signal sequence. Preferably the signal
25 sequence is native (homologous) to the DNA sequence encoding the polypeptide. Alternatively the signal sequence is foreign (heterologous) to the DNA sequence encoding the polypeptide, in which case the signal sequence is preferably endogenous to the host cell in which the DNA sequence is expressed. Examples of suitable signal sequences for yeast host cells are the signal sequences derived from yeast α -factor genes. Similarly, a suitable signal sequence for filamentous fungal host cells is e.g. a
30 signal sequence derived from a filamentous fungal amyloglucosidase (AG) gene, e.g.

the *A.niger glaA* gene. This may be used in combination with the amyloglucosidase (also called (gluco)amylase) promoter itself, as well as in combination with other promoters. Hybrid signal sequences may also be used with the context of the present invention.

- 5 Suitable heterologous secretion leader sequences are those originating from the fungal amyloglucosidase (AG) gene (*glaA* - both 18 and 24 amino acid versions e.g. from *Aspergillus*), the α -factor gene (yeasts e.g. *Saccharomyces* and *Kluyveromyces*) or the α -amylase gene (*Bacillus*).

- 10 The vectors may be transformed or transfected into a suitable host cell as described above to provide for expression of a polypeptide of the invention or for production of vitamin B₁₂. This process may comprise culturing a host cell transformed with an expression vector as described above under conditions to provide for expression by the vector of a coding sequence encoding the polypeptide.

- 15 A further aspect of the invention thus provides host cells transformed or transfected with or comprising a polynucleotide or vector of the invention. Preferably the polynucleotide is carried in a vector for the replication and expression of the polynucleotide. The cells will be chosen to be compatible with the said vector and may for example be prokaryotic (for example bacterial), fungal, yeast or plant cells.

- 20 The invention encompasses processes for the production of the polypeptide of the invention by means of recombinant expression of a DNA sequence encoding the polypeptide. For this purpose the DNA sequence of the invention can be used for gene amplification and/or exchange of expression signals, such as promoters, secretion signal sequences, in order to allow economic production of the polypeptide in a suitable homologous or heterologous host cell. A homologous host cell is herein defined as a host cell which is of the same species or which is a variant within the same species as the species from which the DNA sequence is derived.

- 30 Suitable host cells can be prokaryotic microorganisms such as bacteria, or eukaryotic organisms, for example fungi, such as yeasts or filamentous fungi, or plant cells. In general, yeast cells are preferred over fungal cells because they are easier to manipulate. However, some proteins are either poorly secreted from yeasts, or in

some cases are not processed properly (e.g. hyperglycosylation in yeast). In these instances, a fungal or bacterial host organism can be selected. For production of vitamin B₁₂, a prokaryotic or bacterial host is preferred.

5 The host cell may over-express the polypeptide, and techniques for engineering over-expression are well known³. The host may thus have two or more copies of the encoding polynucleotide (and the vector may thus have two or more copies accordingly).

Bacteria from the genus *Bacillus* are suitable heterologous hosts because of their capability to secrete proteins into the culture medium. Other bacteria suitable as
10 hosts are those from the genera *Streptomyces* and *Pseudomonas*. However, preferably the host is from the same order (e.g. Actinomycetales) or family (*Propionibacteriaceae*) as the bacteria from which the polynucleotides of the invention can be obtained from (*P. freudenreichii*).

A preferred yeast host cell for the expression of the DNA sequence encoding the
15 polypeptide is of the genera *Saccharomyces*, *Kluyveromyces*, *Hansenula*, *Pichia*, *Yarrowia*, and *Schizosaccharomyces*. More preferably a yeast host cell is selected from the group consisting of the species *Saccharomyces cerevisiae*, *Kluyveromyces lactis* (also known as *Kluyveromyces marxianus* var. *lactis*), *Hansenula polymorpha*, *Pichia pastoris*, *Yarrowia lipolytica*, and *Schizosaccharomyces pombe*.

20 Most preferred are, however, (e.g. filamentous) fungal host cells. Preferred filamentous fungal host cells are selected from the group consisting of the genera *Aspergillus*, *Trichoderma*, *Fusarium*, *Disporotrichum*, *Penicillium*, *Acremonium*, *Neurospora*, *Thermoascus*, *Myceliophthora*, *Sporotrichum*, *Thielavia*, and *Talaromyces*. More preferably a filamentous fungal host cell is of the species
25 *Aspergillus oysae*, *Aspergillus sojae*, *Aspergillus nidulans*, or a species from the *Aspergillus niger* Group.²³ These include but are not limited to *Aspergillus niger*, *Aspergillus awamori*, *Aspergillus tubingensis*, *Aspergillus aculeatus*, *Aspergillus foetidus*, *Aspergillus nidulans*, *Aspergillus japonicus*, *Aspergillus oryzae* and *Aspergillus ficuum*, and further consisting of the species *Trichoderma reesei*,
30 *Fusarium graminearum*, *Penicillium chrysogenum*, *Acremonium alabamense*, *Neurospora crassa*, *Myceliophthora thermophilum*, *Sporotrichum cellulophilum*,

Disporotrichum dimorphosporum and *Thielavia terrestris*.

Examples of expression hosts within the scope of the present invention are fungi such as *Aspergillus* species^{31,32} and *Trichoderma* species; bacteria such as *Bacillus* species^{33,34}, e.g. *Bacillus subtilis*, *Bacillus licheniformis*, *Bacillus*
5 *amyloliquefaciens*, *Pseudomonas* species; and yeasts such as *Kluyveromyces* species³⁵ e.g. *Kluyveromyces lactis*,³⁶ and *Saccharomyces* species, e.g. *Saccharomyces cerevisiae*.

Culture of host cells and recombinant production: [cobA or cbiA]

10

The invention also includes cells that have been modified to express the polypeptides of the invention. Preferably the host will have at least two, or multiple, copies of the polynucleotide. Such cells include transient, or preferably stable higher eukaryotic cell lines, such as mammalian cells or insect cells, lower eukaryotic cells,
15 such as yeast and (e.g. filamentous) fungal cells or prokaryotic cells such as bacterial cells (e.g. of the order Actinomycetales).

It is also possible for the proteins of the invention to be transiently expressed in a cell line or on a membrane, such as for example in a baculovirus expression system.

20

According to the present invention, the production of the polypeptide of the invention can be effected by the culturing of microbial expression hosts, which have been transformed with one or more polynucleotides of the present invention, in a conventional nutrient fermentation medium:

25

The recombinant host cells according to the invention may be cultured using procedures known in the art. For each combination of a promoter and a host cell, culture condition are available which are conducive to the expression the DNA
sequence encoding the polypeptide and/or vitamin B₁₂ production. After reaching the desired cell density or titre the culture can be stopped and the polypeptide, or vitamin, is recovered using known procedures.

30

The fermentation medium can comprise a carbon source (e.g. glucose, maltose, molasses, cellulose, β -glucan etc.) and an (inorganic) nitrogen source (e.g. ammonium sulphate, ammonium nitrate, ammonium chloride, etc.) and/or an

(organic) nitrogen source (e.g. yeast extract, malt extract, peptone, etc.). An inorganic nutrient source (e.g. phosphate, magnesium, potassium, zinc, iron, etc.) and/or an inducer (e.g. cellulose, β -glucan, maltose or maltodextrin) may be present.

The selection of the appropriate medium may be based on the choice of
5 expression host and/or based on the regulatory requirements of the expression construct. Such media are known to those skilled in the art. The medium may, if desired, contain additional components favouring the transformed expression hosts over other potentially contaminating microorganisms.

The fermentation can be performed over a period of 0.5-30 days. It may be a
10 batch, continuous or fed-batch process, suitably at a temperature in the range of between 0 and 45°C and, for example, at a pH between 2 and 10. Preferred fermentation conditions are a temperature in the range of between 20 and 37°C and/or a pH between 3 and 9. The appropriate conditions are usually selected based on the choice of the expression host and the protein to be expressed.

15 After fermentation, if necessary, the cells can be removed from the fermentation broth by means of centrifugation or filtration. After fermentation has stopped or after removal of the cells, the polypeptide of the invention may then be recovered and, if desired, purified and isolated by conventional means.

20 D. Uses of the polypeptides in the biosynthetic pathway and to produce vitamin B₁₂
(reaction/enzyme)

A *Amidation ((amide) synthase)*

25 An additional aspect of the invention relates to a process of amidation, or for the preparation of an amine, the process comprising contacting a substrate with a polypeptide of the invention. The process therefore includes amidating a substrate. The polypeptide is preferably a synthase, such as an amide synthase. It may be a polypeptide having the sequence of SEQ ID No. 2, or a variant or fragment thereof,
30 as defined earlier, such as in the first aspect. Alternatively, the polypeptide may be a synthase from a bacterium of the family *Mycobacteriaceae*, for example of the genus

Propionibacterium, in particular the species *Propionibacterium freudenreichii*.

The process may be conducted in the presence of glutamine. The glutamine may be converted to glutamate in the reaction. The polypeptide may be capable of converting a hydroxyl group into an amine, or a carboxyl group (COOH) into a carboxyamide group (CONH₂). The product resulting from the process may therefore be a primary amine.

The process may be repeated, since the polypeptide may amidate the substrate twice, in other words create two (preferably primary) amine groups on (different) substituents of the substrate. Therefore, the process may involve converting a first carboxyl group to a carboxyamide group. The process may be repeated, and a second carboxyl group may be converted to a carboxyamide group as well. In this way, this process may involve amidation twice, for example the creation of two separate (e.g. primary) amines. The (second) amidation preferably takes place at a different substituent (e.g. carboxyl group) on the substrate.

Preferably, the substrate is cobyrinic acid or cobyrinic acid c-amide and/or the product is cobyrinic acid c-amide or cobyrinic a,c-diamide. In this reaction, glutamine can be converted to glutamate. The amount of glutamine added or present may be approximately twice as much as the cobyrinic acid (in other words, glutamine is at a molar concentration of about double the molar concentration of cobyrinic acid). This is because the cobyrinic acid is first amidated to cobyrinic acid c-amide, which acts as an intermediate, and the cobyrinic c-amide is then amidated in a second amidation reaction to give cobyrinic a,c-diamide.

Hence, the polypeptide in this process is preferably a cobyrinic acid a,c-diamide synthase (e.g. cobA or cbiA). The polypeptide may have an activity within EC 6.3.1.-.

B1 (Phospho)transferase (phosphorylation): [cobU]

The invention also relates to a process of phosphorylation, or for the preparation of a phosphate-containing compound, the process comprising contacting a substrate with a polypeptide of the invention. The polypeptide preferably comprises SEQ ID

No. 4, or a variant or fragment thereof, as defined earlier. Alternatively, the polypeptide may be a phosphotransferase from a bacterium of the family *Mycobacteriaceae*, for example of the genus *Propionibacterium*, in particular the species *Propionibacterium freudenreichii*. This process thus comprises
5 phosphorylating (or attaching a phosphate group to) a substrate.

The process may be conducted in the presence of a nucleotidyl (e.g. tri) phosphate, such as ATP. The substrate may comprise a nucleotide, such as one including adenosine.

Preferably the process comprises transfer (from one compound to another, such
10 as to the substrate) of a phosphate moiety, such as phosphorylation of a hydroxyl (OH) group to form a phosphate group ($-\text{PO}_4^-$). The polypeptide may thus act as a phosphotransferase with an alcohol group (such as a hydroxy group) as an acceptor.

Preferably the substrate is adenosyl cobinamide (Formula II) and/or the product is adenosyl cobinamide phosphate (Formula IIA). The process may additionally
15 comprise the conversion of a nucleotide triphosphate to a nucleotide diphosphate (for example, ATP to ADP). The polypeptide may therefore be a cobinamide kinase (e.g. cobU). Preferably, it has an activity within EC 2.7.1.-.

B2 (Nucleotidyl) transferase (nucleotidylation)

20

In this aspect the invention relates to a process of nucleotidylation, or for the preparation of a nucleotidyl-containing compound, the process comprising contacting a substrate with a polypeptide that is a nucleotide transferase. This polypeptide may be the same as that described in B1 above. This is because the (second) enzyme
25 (designated B) has a dual function, and is a bifunctional enzyme.

Thus enzyme B can act as a general transferase, transferring both phosphate groups as well as nucleotidyl groups. It can therefore act as both a phosphotransferase (B1) and as a nucleotidyl transferase (B2).

The second function, designated B2, relates to the activity of the enzyme as a
30 nucleotidyl-containing transferase.

Thus, preferably the process involved nucleotidylating a substrate, such as

guanidylating (a substrate). Preferably, the substrate will comprise at least one phosphate group. Suitably, the polypeptide is able to nucleotidylate a phosphate group.

5 The process may take place in the presence of a nucleosyl (e.g. tri) phosphate, for example GTP). Thus in this process the polypeptide preferably catalyses the guanidylation of a phosphate group.

Preferably the substrate is adenosyl cobinamide phosphate (Formula IIA) and/or the product is adenosyl-GDP-cobinamide (Formula IIB). Thus the enzyme can catalyse the formation of a nucleotidyl-containing compound, such as adenosyl-GDP-cobinamide, from a substrate such as adenosyl cobinamide phosphate. The polypeptide may thus be a (nucleotidyl) transferase, or has an activity within EC 2.7.7.-. Other preferred features of the polypeptide are as described in the previous section concerning phosphotransferase activity (B1).

15 C Arylation (aryl transferase) or ribazole addition: [cobS]

This process of the invention comprises arylation, or the preparation of an aryl-containing compound, the process comprising contacting a substrate with a polypeptide of the invention, preferably an (e.g. aryl) transferase. Preferably the polypeptide comprises SEQ ID No. 6, or a variant or fragment thereof as previously defined. Alternatively, the polypeptide may be an aryltransferase from a bacterium of the family *Mycobacteriaceae*, for example of the genus *Propionibacterium*, in particular the species *Propionibacterium freudenreichii*. This process thus comprises arylating a substrate.

25 The aryl moiety (e.g. in ribazole, such as is transferred during a reaction) may comprises an aromatic ring system. The aryl moiety may comprise one or two aromatic rings. The ring system may be substituted by from one to four C₁₋₈ alkyl groups. The aryl moiety may comprise none, one or two heteroatoms, for example one or two nitrogen atoms. Preferably the aryl moiety comprises a benzimidazole ring. The process therefore may comprise the preparation of a (e.g. dimethyl) benzimidazole (DMB)-containing compound.

The aryl group may be bonded or joined to a (central) metal, e.g. cobalt, atom. Alternatively or in addition, the aryl group may be bonded to a carbon atom, such as in a ribose group. Preferably the aryl moiety is bonded to both cobalt atom and a ribose group (in the product of the reaction, namely the resulting benzimidazole-
5 containing compound).

The process may take place in the presence of a ribazole, such as α -ribazole. This may be present in an approximately equimolar amount to the substrate. The reaction may comprise alpha-ribazole addition (to the substrate).

Preferably the substrate is adenosyl-GDP-cobamide. The product of the
10 reaction, the aryl-containing compound, is preferably adenosyl-5,6-dimethyl benzimidazolyl cobamide (Formula IIC). In the process of the reaction, the ribazole may be converted to GMP.

Preferably the polypeptide used in this process is a cobalamin (5'-phosphate) synthase (e.g. cobS). The polypeptide may have the activity EC 2.7.8.-.
15

D Adenosylation (adenosyl transferase)

This process of the invention relates to adenosylation, or the preparation of an adenosine-containing compound, the process comprising contacting a substrate with
20 a polypeptide of the invention, preferably a transferase, such as an adenosyl transferase. The polypeptide preferably comprises SEQ ID No. 7, of fragment or variant thereof, as previously defined. Alternatively, the polypeptide may be a transferase from a bacterium of the family *Mycobacteriaceae*, for example of the genus *Propionibacterium*, in particular the species *Propionibacterium freudenreichii*.
25 This process thus comprises adenosylating a substrate.

This process may therefore comprise the transfer of adenosine, preferably to the substrate. Preferably the adenosine becomes bound to a metal atom, such as a transition metal (such as of the first series), for example cobalt.

The substrate (and/or product) may be an amide, such as a diamide. Preferably
30 the substrate is cobyrinic acid a, c diamide and/or the product is adenosyl cobyrinic acid a, c diamide.

The process may take place in the presence of a nucleosyl (e.g. tri) phosphate, such as ATP. It may also take place in the presence of adenosine. Preferably, both the adenosine and the nucleosyl phosphate are present in approximately equimolar amounts to the substrate. The nucleosyl triphosphate may be converted to a

5 nucleosyl diphosphate.

Preferably the polypeptide in this process is an adenosyl transferase. It may have an activity within EC 2.5.1.7. Preferably, the polypeptide is a transferase that is capable of transferring alkyl or aryl groups, other than methyl groups. Polypeptides that methylate, or cause methylation, may thus be excluded.

10

Substrates (or products of catalysed reaction): vitamin B₁₂ intermediates and/or precursors

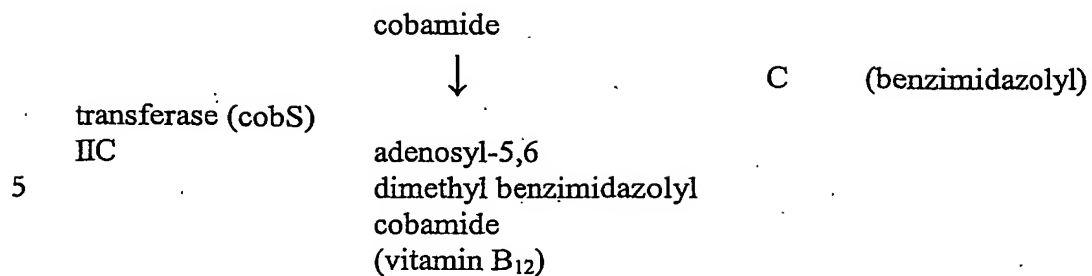
The substrate and/or product preferably comprises a corrin core or ring
15 system. Preferably, it comprises an aryl ring system, with up to four rings (which may be the same or different). Preferably however there are four rings, and they are the same. Each ring may contain one or two heteroatoms, for example one nitrogen atom. The ring may be pentagonal. Thus preferred rings are pyrrole, and therefore the ring system preferably comprises a tetrapyrrole system. Preferably two of the
20 pyrrole rings are joined to each other, and the other two pyrrole rings are joined by a bridge, such as by methene units.

The ring system may comprise a metal atom, for example at its core. This metal may be a transition metal, for example of the first series (group VIII). It may be of period four. Preferably the metal is cobalt, and this may be a single central
25 cobalt atom. The ring system may have attached to it an amide, phosphate, guanidyl or adenosyl moiety or group. In addition to the metal atom and ring system, there may be a fifth, and optionally even a sixth substituent, for example bound to the metal. The or each substituent may be above and/or below the plane of the ring, as applicable. Where appropriate (and in the case of vitamin B₁₂), one of these
30 substituents may comprise a nucleosyl group, such as a dioxy nucleosyl, preferably 5'-dioxy adenosine. Another substituent may be an aryl group as defined above in

section C concerning arylation. This substituent may therefore preferably comprise dimethoxybenzimidazole. In substrates employed in the invention, one or both substituents may be present, and so that the cobalt atom may have, as fifth and sixth substituents, a 5'-dioxy adenosine and a dimethoxy benzimidazole group. The same preferred features apply to the product of the reaction as they do to the substrate.

A list of the substrates and products of the five catalysed reactions, or biosynthetic steps, is provided below after the role of the enzymes in the vitamin B₁₂ biosynthetic pathway.

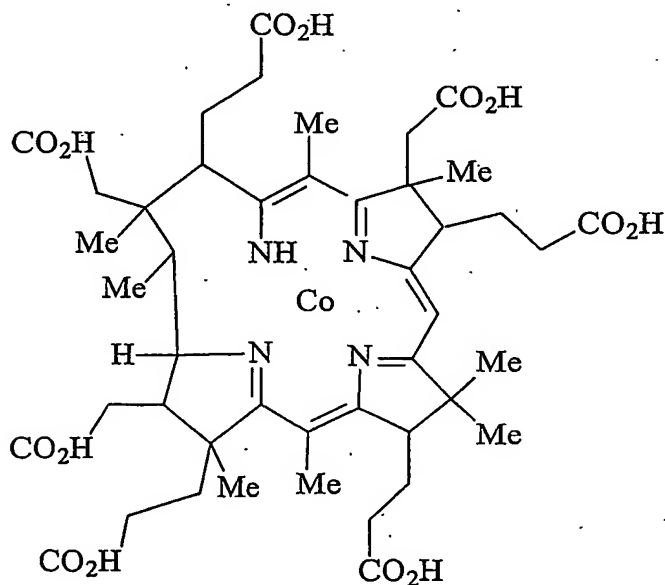
10	A (cobA)	B (cobU)	C (cobS)
	Uro III → → → → Cobinamide → cobinamide-P → Vitamin B ₁₂		
	Formula	Common name	Reaction / Biosynthetic step:
15	Enzyme		
	I	cobyritic acid	
		↓	
	IA	cobyritic acid c-amide	A amide synthase
20		↓	
	IB	cobyritic acid a,c-diamide	A amide synthase
		↓	
	IC	adenosyl cobyritic acid-a,c-diamide	D (adenosyl) transferase
25			
	II	adenosyl cobinamide	
30		↓	
	(cobU) IIA	adenosyl cobinamide phosphate	B1 (phospho) transferase
		↓	
35	(cobU) IIB	adenosyl-GDP	B2 (nucleotidyl) transferase



Preferred intermediates or precursors include cobinamide, cobinamide-P
 (phosphate) or
 compounds of Formula 1A, 1B, II, IIA, IIB or IIC.

Hence the process of the present invention may comprise one or more of the
 following process steps (illustrated using chemical formulae), namely:

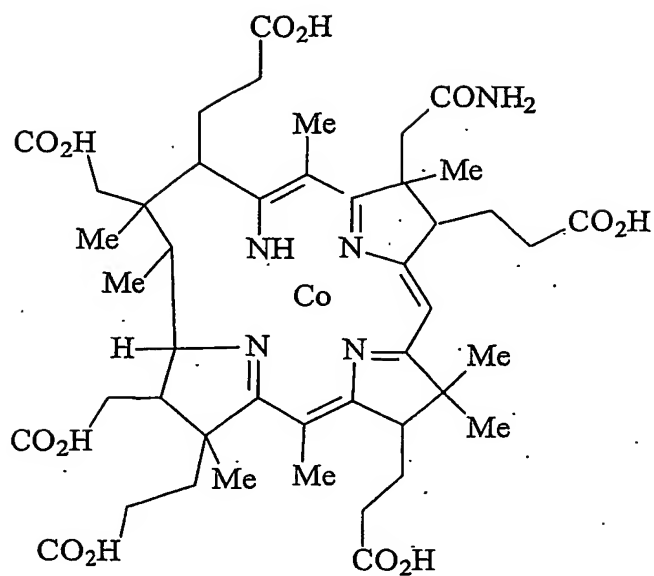
Formula I



↓ (amide) synthase (A)

(SEQ ID No. 2, or variants, etc thereof)

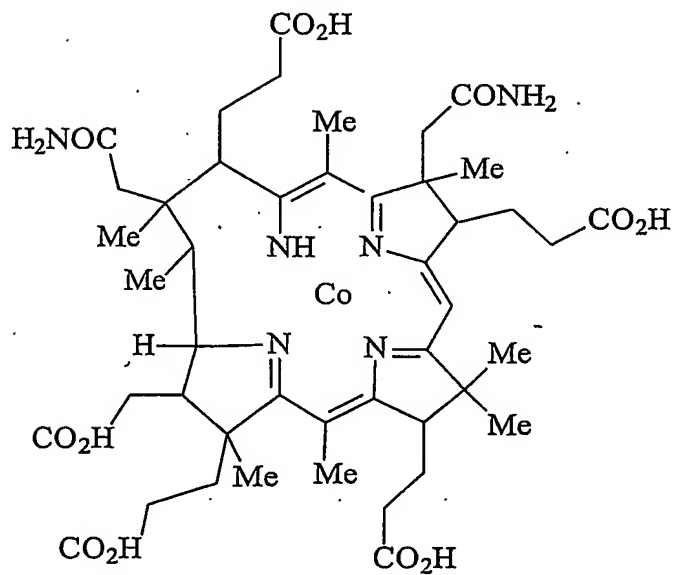
Formula IA



↓ (amide) synthase (A)

(SEQ ID No. 2, or variants, etc thereof)

Formula IB

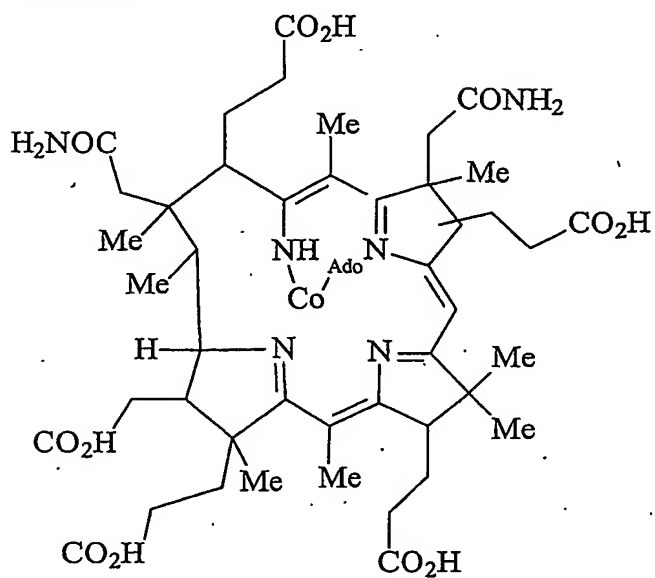


40

↓ adenosyl transferase (D)

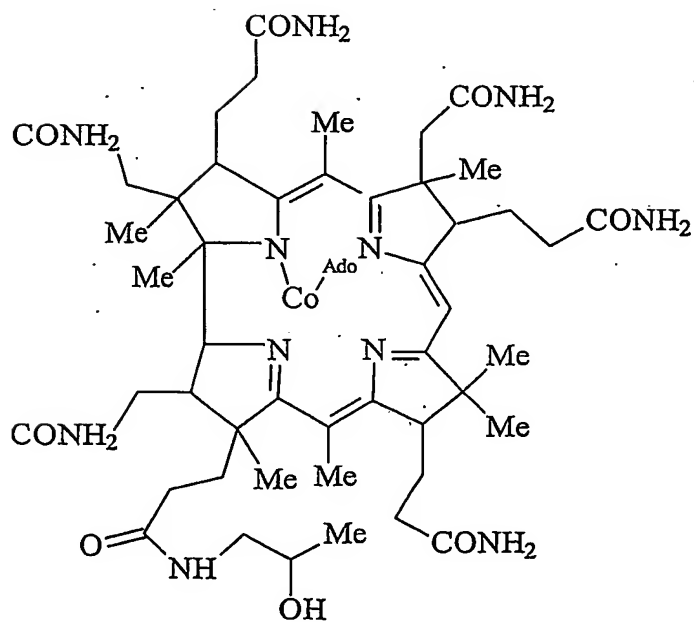
(SEQ ID No. 8, or variants, etc thereof)

Formula IC



5

Formula II



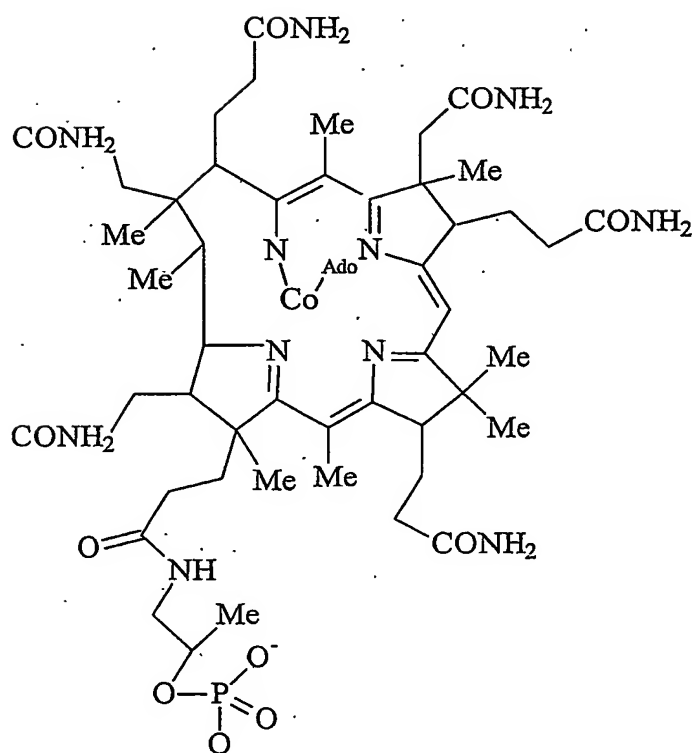
41

↓ (phospho) transferase (B1)

(SEQ ID No. 4, or variants, etc. thereof)

5

Formula IIA

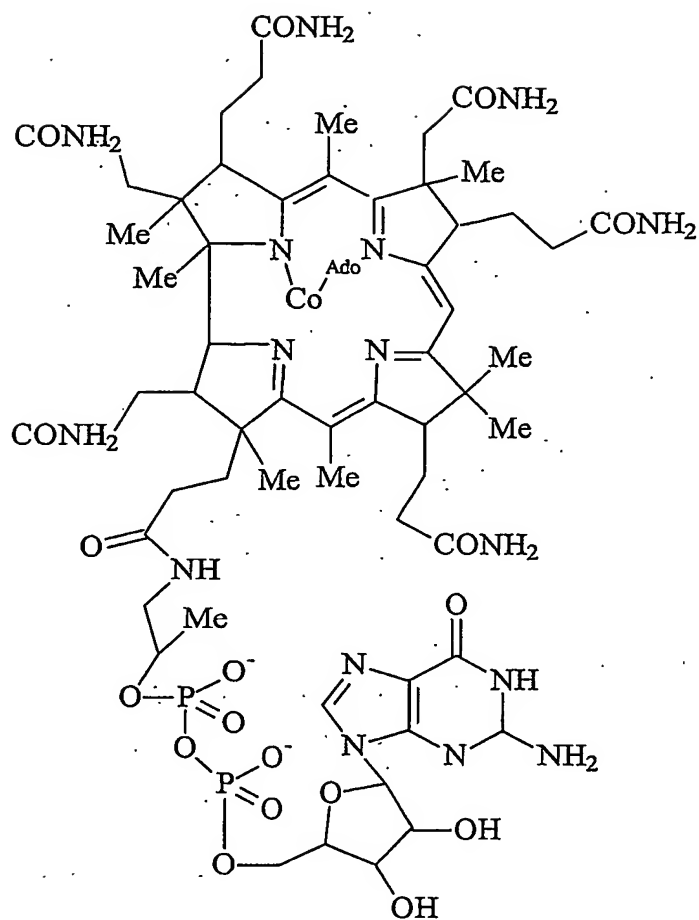


10

↓ (nucleotidyl) transferase (B2)

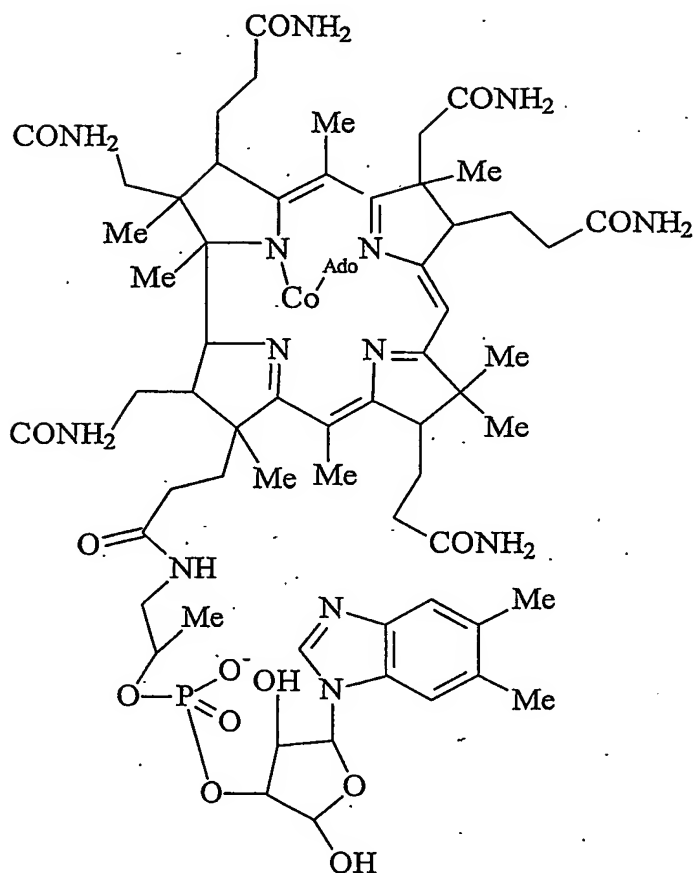
(SEQ ID No. 4, or variants, etc. thereof)

Formula IIB



↓ (benzimidazolyl) transferase (C)

(SEQ ID No. 6; or variants, etc. thereof)

Formula IIC (includes vitamin B₁₂)E. Industrial preparation of vitamin B₁₂

5

As described above, the polypeptides of the invention may be used to perform one or more biosynthetic steps in the vitamin B₁₂ preparatory pathway. The polypeptide may be contacted with the appropriate substrate and the reaction allowed to take place. This may be allowed to occur when the polypeptide is outside (e.g. host) cell, in other words the polypeptide is mixed with the substrate, for example *in vitro*. However, it is considerably more effective to use a host cell of the invention, comprising one or more of the polypeptides of the invention, in order to either

10

perform one or more particularly desired steps in the vitamin B₁₂ biosynthetic pathway or, preferably, use the host cell to produce vitamin B₁₂ (or an intermediate or precursor thereof).

5 The invention therefore also relates to a process for preparing vitamin B₁₂ (or a precursor thereof), the process comprising culturing one or more host cells of the invention under conditions that allow the cells to biosynthesise (and hence produce) vitamin B₁₂ (or the precursor).

10 This process may take place in a fermenter. The fermenter may be equipped with agitation means, for example a stirrer. The fermenter vessel may also be equipped with aeration means, such as a means of causing an oxygen-containing gas to be contacted with a liquid in the fermenter. The liquid will usually be the culture broth, consisting of an aqueous suspension of the cells. Fermentation may then be allowed to take place. The fermenter may have a minimum volume of 10, 50, 100 or
15 1,000 litres.

 The cells may be supplied with one or more carbon and/or nitrogen sources, before fermentation begins, at the start of fermentation, or continuously or continually during fermentation.

20 At the end of the fermentation, the supply of the carbon or nitrogen sources is stopped, or one or more of these sources may be used up. Each of the carbon and/or nitrogen sources may be a complex source or an organic or inorganic compound.

 The cells may then be removed from the fermenter. Before or after this, water (or an aqueous liquid) may be removed from the cells. The cells may then be heated or pasteurised, in order to kill them.

25 Methods of extracting or isolating vitamin B₁₂ from microbial cells are well known⁴⁵. For example (in order to be able to obtain the vitamin B₁₂) it is preferred that the host cells, having produced the vitamin, are broken, or at least partially opened, so that at least part of the soluble content of the cells (comprising vitamin B₁₂) is released into a liquid, for example a liquid in which the cells are contained.
30 One can then separate the open or broken cells, or resulting cell debris, from the liquid (comprising the vitamin B₁₂). The microbial cells (containing vitamin B₁₂)

can thus be treated so as to cause disruption of the cell membranes. Suitable treatments for opening the cells include heat treatment, such as pasteurisation, heating in an autoclave, treatment with bacteriolytic enzymes (such as lysozyme), and/or mechanical disruption cells (grinding, or the use of shear forces), or treatment
5 of chemicals (to cause cell lysis, for example the use of detergents or organic solvents).

The process of lysis or other membrane disruption can produce a lysate, which can then be separated into solid and liquid phases. The solid phase of the lysate, comprising the cell debris, can then be separated (from the liquid containing
10 vitamin B12). A number of suitable solid-liquid separation techniques are available, including centrifugation and/or filtration. Preferably however, solid liquid separation is performed using ultrafiltration.

Preferably, the open/broken microbial cells are washed, and the washings are then combined or added to the (vitamin B12 containing) liquid, separated from cell
15 debris. Suitably washing comprises diafiltration, suitably with ion-free water. The (vitamin B12) containing diafiltrate can then be combined with the (vitamin B12 containing) liquid phase.

The vitamin B12 containing liquid(s) may then be subjected to drying, for example spray-drying, fluid-bed drying, freeze drying or drying in a vacuum.

20 Preferably, cells producing vitamin B12 are washed prior to opening (lysis), since this may increase the vitamin B12 concentration on dry matter by removing medium components. This can be performed using diafiltration, preferably using ion-free water.

Preferred features and characteristics of one aspect of the invention are
25 applicable to another aspect *mutatis mutandis*.

The invention will now be described with reference to the following Examples which are intended to be illustrative only and not limiting.

EXAMPLE 1

30

Propionibacterium vectors

Two strains (*P. freudenreichii* LMG16545 and *P. freudenreichii* LMG16546) were used which both show an identical plasmid profile of 2 plasmids. One plasmid was large (size not determined) and the other was smaller, more abundantly present and had a size of 3.6 kb. These 3.6 kb plasmids from LMG16545 and LMG16546 were chosen for further use in vectors. Plasmids from these vectors already are described.²⁹ Expression systems in *Propionibacteria* are known in the art³⁰.

Construction of *E. coli/Propionibacterium* shuttle vectors

A 1.7 kb *Acc*65I fragment from the *Saccharopolyspora erythraea* NRRL2338 erythromycin biosynthesis cluster and containing the erythromycin resistance conferring gene^{37, 38} was inserted into *Acc*65I linearized pBR322ΔI²⁹. Then the newly derived construct, named pBRES, was linearized with *Eco*RV and ligated to p545 DNA that had been digested with *Bsa*BI. *E. coli* transformants were found to harbor a vector with the correct insert, in both orientations. The resulting plasmid vectors were named pBRESP36B1 and pBRESP36B2 (see Figs. 2a and 2b²⁹).

Plasmid vector constructs were also obtained with p545 DNA linearized in an other restriction site situated outside the putative replication region, namely *Alw*NI. For this construction the pBRES vector had to be provided with a suitable cloning site. An adaptor was designed and annealing of the required oligo's created a double stranded DNA fragment with *Acc*65I and *Bgl*II cohesive ends respectively, which moreover contains an internal *Sfi*I restriction site, that provides ends compatible to the *Alw*NI digested p545 plasmid. This adaptor was cloned in pBRES between the *Bgl*II and the proximal *Acc*65I site. The pBRES-*Sfi*I vector thus obtained was subsequently digested by *Sfi*I and ligated to *Alw*NI digested p545. Transformation of *E. coli* yielded transformants with the correct vector as confirmed by restriction enzyme analysis. The vector obtained was named pBRESP36A²⁹.

Transformation of *Propionibacterium* with *E. coli/Propionibacterium* shuttle vectors

Transformation of *Propionibacterium freudenreichii* strain ATCC6207 with pBRESP36B1 will be described.

The bacterial cells are cultivated in SLB (sodium lactate broth³⁹ at 30°C to a

stationary growth phase, and subsequently diluted 1:50 in fresh SLB. After incubation at 30°C for around 20 hours, cells (now in the exponential growth phase) were harvested and washed extensively in cold 0.5M sucrose. Subsequently cells were washed once in the electroporation buffer, consisting of 0.5M sucrose, buffered
5 by 1mM potassium acetate, pH5.5, and finally resuspended in this electroporation buffer in about 1/100 of the original culture volume. Cells were kept on ice during the whole procedure.

For the electroporation (apparatus from BIORAD), 80 - 100 µl of cell suspension was mixed with ± 1 µg of DNA (or smaller amounts), in a cooled 1 or 2
10 mm electroporation cuvette, and an electric pulse delivered. Optimal pulse conditions were found to be 25kV/cm at 200 Ω resistance and 25µF capacitance. However, lower and higher voltages (also at 100Ω) also yield transformants.

Immediately after the pulse, 900 µl cold SLB containing 0.5 M sucrose was added to the pulsed cell suspension and these are subsequently incubated for 2.5 to 3
15 hours at 30°C before plating appropriate dilutions on SLB/agar plates containing 0.5 M sucrose and 10µg/ml erythromycin. After a 5 to 7 day incubation period at 30°C under anaerobic conditions, transformants were detected.

DNA isolated from *E. coli* DH5α (Promega) yielded a transformation efficiency of 20 - 30 transformants per µg DNA. A 10-100 fold higher efficiency is achieved
20 when DNA is isolated from *E. coli* JM110 (dam⁻, dcm⁻ strain). *E. coli* transformation was done according to BIORAD's instructions.

Transformants contained the authentic vectors, indistinguishable from the original plasmid DNA used for transformation of ATCC6207. This was shown by restriction enzyme analysis of plasmid DNA isolated from the transformants by the
25 small scale plasmid DNA isolation procedure referred to before.

Vectors were exclusively present as autonomously replicating plasmids. Southern blot hybridization⁴⁰ with total DNA isolates showed that chromosomal DNA did not hybridise to the vector DNA used as a probe, indicating that no chromosomal integration of plasmid DNA occurred.

30 Transformation was also successful with vectors pBRESP36B2 and pBRESP36A, indicating that functionality of the vector was independent of the

orientation of p545 or the cloning site used. Also in this case the authenticity of the vectors was confirmed.

Moreover, transformation of *P. freudenreichii* strain ATCC6207 with DNA isolated from a *Propionibacterium* transformant resulted in a 10^5 - 10^6 fold increased transformation efficiency as compared to that obtained with DNA isolated from *E. coli* DH5 α .

Transformation of another *P. freudenreichii* strain, LMG16545 (the same strain from which the p545 plasmid was obtained), resulted in a transformation efficiency comparable to that of the ATCC strain.

The transformations we repeated using each of SEQ ID Nos. 1, 3, 5 and 7 operably linked to appropriate transcription and translation start signals in the shuttle vector.

EXAMPLE 2

Construction of plasmid vector containing the amide synthase (A) gene

The construction and application of a plasmid vector to increase the level of vitamin B₁₂ (cobalamine) synthesis in *P. freudenreichii* strain ATCC6207 will be described.

For construction of gene overexpression plasmids use was made of the 16S rRNA promoter from *P. freudenreichii*. One of the strategies to test promoter bearing fragments is the use of a promoter-probe vector. The reporter gene used to monitor promoter activity encodes an easily detectable enzymatic activity that is not present in the wild type strain. The *cat* (chloramphenicol acetyl transferase) gene from pACYC184^{17, 42} was used for construction of a promoter-probe vector. In order to analyse the activity of the 16S rRNA promoter this promoter was placed upstream of the *cat* gene.

For construction of the promoter-probe vector, the promoterless *cat* gene was cloned in the *E. coli* / *Propionibacterium* shuttle vector pBRESP36B2²⁹ by PCR, resulting in vector pB2/PoCAT. The upstream PCR primer included the sequence 5'-

GGGATCCTCTAGAGCATGCAAGCTTCTCGAGAATCGATAGATCTCTAAGG
AAGCTAAAATG-3', in which the last three nucleotides indicate the start codon of
the *cat* gene. This synthetically derived sequence includes a multi-cloning site (MCS)
containing the restriction sites BamHI, XbaI, HindIII, SphI, XhoI, ClaI and BglII. The
5 downstream PCR primer included a BamHI restriction site. After PCR amplification
the *cat* gene was cloned as a BamHI fragment in the BglII site of the vector (BamHI
and BglII sites not restored). Two orientations of the *cat* gene were obtained. The
orientation in which the *cat* gene has the same orientation as the beta lactamase gene
in the pBR322 segment was used in further experiments.

10 On the basis of the sequence of 16S rRNA from *Propionibacterium*
freudenreichii ATCC6207 (GenBank accession number X53217) an appropriate
restriction enzyme was chosen (HindIII) and appropriate PCR primers were designed
that enabled the amplification of an approximately 3 kb region encompassing the
promoter by inverse PCR². From the PCR fragment a 0.6 kb SphI-HindIII fragment
15 directly upstream of the 16S rRNA coding sequence was isolated. This fragment was
ligated in pB2/PoCAT digested with the same enzymes resulting in a plasmid named
pB2/PrRNACAT. After transformation of *E. coli* chloramphenicol resistant
transformants were obtained. After transformation *P. freudenreichii* strain ATCC
6207 colonies were only obtained on erythromycin containing plates, not on
20 chloramphenicol containing plates. However, when streaked on chloramphenicol
containing plates transformants containing pB2/PrRNACAT grew whereas
ATCC6207 cells containing pB2/PoCAT did not grow, thus indicating the
functionality of the 16S rRNA in *P. freudenreichii*.

An expression vector containing the 16S rRNA promoter but lacking the *cat*
25 gene was constructed by ligation of the approx. 700 bp BamHI - BglII fragment from
vector

pB2/PrRNACAT, containing the 16S ribosomal RNA promoter from *P.*
freudenreichii, into the unique BglII of pBRESP36B2. Both possible orientations of
the promoter element in the vector were obtained. In case transcription of genes
30 expressed by this ribosomal promoter was not properly terminated, readthrough may
obstruct transcription of the two replication genes of the *Propionibacterium* replicon

if the ribosomal promoter is oriented towards these replication genes. Therefore the vector in which the promoter was cloned in the opposite orientation, pBRESP36B2p16SH, was used in further experiments. A unique BglII site downstream from the promoter was used for cloning of the expression library.

5

EXAMPLE 3

The complete coding sequence of adenosyl transferase (D, SEQ ID No. 7) was generated by PCR from appropriate primers. The upstream PCR primer included a 5' extension including a BglII restriction site and a ribosome binding site upstream of the start codon of the gene. The downstream PCR primer included a 5' extension including a BglII restriction site. After digestion of the amplified fragment with BglII the fragment was ligated in the vector pBRESP36B2p16SH and digested with BglII and dephosphorylated to remove 5' phosphate groups. After transformation of *E. coli* ampicillin resistant colonies were obtained. Both orientations of the cloned fragment relative to the vector were observed. However, only the construct in which the 16S rRNA promoter is located directly upstream of the ribosome binding site allows the expression of the adenosyl transferase gene.

The latter construct was transformed to *P. freudenreichii* ATCC6207 as described before. In the transformants the level of vitamin B₁₂ synthesis was determined as follows. Frozen cultures of *Propionibacterium* transformants, as well as a control strain containing only the vector plasmid pBRESP36B2, were inoculated in 100 ml flasks containing 50 ml of BHI (Brain Heart Infusion) medium (Difco) and incubated for 72 hrs at 28°C without shaking. From this preculture 4 ml were transferred to 200 ml of production medium consisting of Difco yeast extract 15 g/l, Na lactate 30 g/l, KH₂PO₄ 0.5g/l, MnSO₄ 0.01 g/l, and CoCl₂ 0.005 g/l in a 500 ml shake flask and incubated at 28°C for 56 hrs without shaking, followed by 48 hrs in a New Brunswick rotary shaker at 200 rpm.

Vitamin B₁₂ titres were measured using HPLC⁴³ and showed a higher vitamin B₁₂ production than the control strain.

The process was repeated for each of the other three genes namely:

- A: SEQ ID No. 1, cobyrinic acid -a, c-diamide synthase;
B: SEQ ID No. 3, cobinamide kinase gene; and
C: SEQ ID No. 5, cobalamin (5'-phosphate) synthase.

The four genes (A, B, C, and D) were then combined in one operon to further
5 increase vitamin
B₁₂ production.

The resulting transformed cells (the *P. freudenreichii* ATCC 6207) was
cultured in a fermenter using a known technique⁴⁴. In order to kill the cells, and to
cause lysis, the broth was pasteurised at 65°C for thirty minutes. The broth was then
10 subjected to ultrafiltration, and a pink coloured filtrate was obtained, containing
vitamin B₁₂. The heat had caused lysis of the cells, and therefore release of the
intracellular vitamin B₁₂ in the medium.

15 EXAMPLE 4

Construction of expression vectors for *Propionibacteria* and its use in expression of enzymes of the invention and production of vitamin B₁₂ in *Propionibacterium freudenreichii*

20 Apart from the expression systems described above^{29,30}, two other expression
systems are known in the art that can be used to express, in multiple copies if desired,
genes encoding the novel enzymes of the invention. The first is pRG01, a plasmid
from *Propionibacterium acidipropionici*²⁷. This was used to create a shuttle vector
pPK705. Such a vector was used to carry successfully the cobyrinic acid a,c-diamide
25 synthase (A) enzyme, and thus transform *Propionibacterium freudenreichii*
subspecies *shermanii*.

The other suitable expression system that was employed used the known shuttle
vector pPK705²⁸, which is able to shuttle between *E. coli* and *Propionibacterium*.
This allowed the construction of expression vectors for *Propionibacteria*, and the
30 incorporation of enzyme B, cobinamide kinase, into *Freudenreichii* subspecies
shermanii. This was achieved by using the cobinamide kinase gene, SEQ ID No. 3
of the invention.

COMPARATIVE EXAMPLE 5Shake flask fermentation of *Propionibacterium* containing an empty plasmid vector

5 The transformation of *P. freudenreichii* strain ATCC6207 with the *E. coli* / *Propionibacterium* shuttle vector pBRESP36B2p16SH has been described in Example 2. Transformants were selected on erythromycin containing agar plates as described before and transformants containing the authentic plasmid were selected by restriction analysis.

10 Transformants were grown in brown-glass penicillin flasks containing 50 ml of anaerobic M1 culture medium prepared as follows. 900 grams of M1 culture medium was prepared by adding 90 ml of part B to 810 grams of part A under anaerobic conditions. Erythromycin was added to a final concentration of 10 milligram / liter.

15 810 grams of part A consisted of 52.2 g MES, 12.51 g Difco Yeast Extract and 0.68 g methionine dissolved in water. The pH was adjusted to 7 with ammonium hydroxide and the oxygen was removed by degassing and flushing with nitrogen. Finally part A was sterilized in an autoclave.

20 90 ml of part B consisted of 1.5 ml trace element solution, 1.5 ml filter sterilized CoCl₂·6H₂O (5g/kg), 0.9 ml filter sterilized Ca-pantothenate (20g/kg) and 86.1 ml degassed sterile glucose (440 g/kg).

1 kg of trace elements solution contained 10g citric acid, 10 ml H₂SO₄, 1.2 g MnSO₄·H₂O, 300 g MgSO₄·7H₂O, 6 g FeSO₄·7H₂O and 2.4 g ZnSO₄·7H₂O in water. The solution was filter sterilized.

25 The 50 ml M1 medium was inoculated under anaerobic conditions with 5 ml of an pre-culture that consisted of a selected pBRESP36B2p16SH transformant that had been grown anaerobically in M1 culture medium for 3 days at 30 °C with gentle shaking until an OD₆₃₀ of approximately 10.

30 The 50 ml culture was incubated for 40 hours at 30°C without shaking. At this time-point a sample was taken from the culture and the OD₆₃₀ determined (=OD-ana). The pH of the medium was corrected through the anaerobic addition of NaOH

(10%). The number of microliters of NaOH (10%) needed were calculated using the formula: number of microlitres NaOH (10%) = $(19.67 \times \text{OD-ana}) - 60$. As a next step 0.5 ml of DBI (0.5 g/kg) was anaerobically added. Following these additions the culture was incubated for another 24 hrs at 30°C with shaking at 200 rpm. The OD630 was determined again (=OD-end).

10 ml samples of the cultures were collected. The cells were harvested by centrifugation and the pellet was washed once with 1 volume sterile demineralised water. The pellet was stored at -20°C. Thawed samples were heated to 80°C in a 2% KCN containing buffer to open the cells and to stabilise vitamin B12 and precursors.

Vitamin B12 titers as well as adenosyl cobinamide and adenosyl cobinamide phosphate precursor levels were measured using HPLC using appropriate standards (43). The identity of the adenosyl cobinamide and adenosyl cobinamide phosphate precursor peaks was established on the basis of mass spectrometry data.

The amount of vitamin B12 produced per ml culture of pBRESP36B2p16SH transformants was divided by OD-end. The resulting ratio obtained for pBRESP36B2p16SH transformants was used as a reference for the ratios obtained for *P. freudenreichii* transformants containing other plasmids (e.g. next example). Similar ratios were calculated for the amounts of adenosyl cobinamide and adenosyl cobinamide phosphate produced (amount per ml culture per OD-end). These ratios were used as reference for the ratios obtained for respectively adenosyl cobinamide and adenosyl cobinamide phosphate in *P. freudenreichii* transformants containing other plasmids.

EXAMPLE 6

Construction of an expression vector for the *Propionibacterium freudenreichii* cobA/cbiA gene and its use for the production of vitamin B12 and its precursors in *Propionibacterium freudenreichii*

A *Propionibacterium freudenreichii* gene coding for glutamate 1-semialdehyde aminomutase named hemL⁴⁶ (EMBL accession number D12643) was

used. The coding region of this gene is followed by the transcriptional hemL terminator. A DNA fragment containing the coding region of the hemL gene and the hemL terminator was cloned in pBRESP36B2p16SH as described in the following steps. The DNA fragment was amplified by PCR from chromosomal DNA isolated
5 from *P. freudenreichii* ATCC6207 using the following primers: 5'-CAgTAgATCTCgACAAGgAggAACCCAtgAg-3' and 5'-CgTAAgATCTCagTTTCggACATggCagTg-3'. Both primers contain a BglII restriction site. The PCR fragment obtained was ligated into the vector pCR-Blunt II-TOPO (Invitrogen) to create pGBPHEL-1. After transformation of *E. coli* kanamycin
10 resistant colonies were obtained. The constructs thus obtained were verified by restriction analysis and the DNA sequence of the fragment was verified through sequence analysis.

pGBPHEL-1 was subsequently digested with BglII and the fragment containing the hemL gene was isolated and ligated into the vector
15 pBRESP36B2p16SH which first had been digested with BglII and dephosphorylated. After transformation of *E. coli* ampicillin resistant colonies were obtained. A construct in which the hemL gene is transcribed from the 16S promoter was selected by restriction analysis. The resulting construct was called pGBP01HEL-1.

A *P. freudenreichii* gene coding for uroporphyrinogen III methyltransferase
20 named cobA (EMBL accession number U13043) was used⁴⁷. In order to over-express the cobA gene in *Propionibacterium* the gene was placed behind the 16S rRNA promoter and in front of the hemL terminator present in pGBP01HEL-1 as described in the following steps. A DNA fragment containing the cobA coding region was amplified by PCR from chromosomal DNA isolated from *P. freudenreichii*
25 ATCC6207 using the following primers: 5'- CACCACCAACATCgATgAggAAAC-3' and 5'- TCCAATTgggACTCagTggTCgCTg-3'. The first primer contains a ClaI and the second primer contains a MfeI restriction site. The PCR fragment obtained was ligated into the vector pCR-Blunt II-TOPO (Invitrogen). After transformation of *E. coli* kanamycin resistant colonies were obtained. The DNA sequence of the cloned
30 fragment was verified through sequence analysis. The resulting construct was named pGBPCOB-1.

pGBPCOB-1 was subsequently digested with ClaI and MfeI and the fragment containing the cobA gene was isolated and ligated into the large vector fragment isolated from pGBP01HEL-1 digested with the same enzymes. After transformation of *E. coli* ampicillin resistant colonies were obtained. Plasmid DNA isolated from transformants was verified by restriction analysis and the resulting construct was called pGBP02COB-1.

pGBP02COB-1 was transformed to *E. coli* JM101 and plasmid DNA isolated from the obtained transformants was verified by restriction analysis. The plasmid DNA was subsequently transformed to *P. freudenreichii* ATCC6207 as described before. Transformants were selected on erythromycin containing agar plates as described before. The plasmid DNA isolated from the *P. freudenreichii* transformants was verified by restriction analysis.

Shake flask fermentations of pGBP02COB-1 transformants using M1 culture medium and the determination of vitamin B12, adenosyl cobinamide and adenosyl cobinamide phosphate levels were performed as described in the previous Example.

Ratios of the amount of vitB12, adenosyl cobinamide and adenosyl cobinamide phosphate produced per ml culture OD-end were calculated as described in the previous example. The ratio obtained for vitB12 was 6% lower than that obtained for pBRESP36B2p16SH transformants in the previous example. However, the ratio obtained for adenosyl cobinamide was 227% higher than that obtained for pBRESP36B2p16SH transformants and the ratio obtained for adenosyl cobinamide phosphate was 200% higher than that obtained for pBRESP36B2p16SH transformants. Thus there was a 2-fold increase in cobinamide and 3-fold increase in cobinamide P over the empty plasmid (Comparative Example 5).

EXAMPLE 7

Construction of an expression vector for the *Propionibacterium freudenreichii* gene encoding protein B (cobU) and its use for the production of vitamin B12 and its precursors in *Propionibacterium freudenreichii*

In order to over-express the gene coding enzyme B (SEQ ID No.3) in *Propionibacterium* the gene was placed behind the 16S rRNA promoter and in front of the hemL terminator present in pGBP01HEL-1 as described in the following steps. A DNA fragment containing the coding region for B was amplified by PCR from
5 chromosomal DNA isolated from *P. freudenreichii* ATCC6207 using the following primers: 5'-CTgATATCAATTggAggACATCAGATgACCCgCATCgTC-3' and 5'-CTgAATTCggCCACgTCAGATCgCgTCC-3'. The first primer contains an EcoRV and a MfeI restriction site and the second primer contains an EcoRI restriction site. The PCR fragment obtained was ligated into the vector pCR-Blunt II-TOPO
10 (Invitrogen). After transformation of *E. coli* kanamycin resistant colonies were obtained. The DNA sequence of the cloned fragment was verified through sequence analysis. The resulting construct was named pGBPCOU-1.

pGBPCOU-1 was subsequently digested with EcoRV and EcoRI and the fragment containing the gene coding for B was isolated and ligated into the large
15 vector fragment isolated from pGBP01HEL-1 which was first digested with ClaI, than blunted with Klenow enzyme, and finally digested with MfeI. After transformation of *E. coli* ampicillin resistant colonies were obtained. Plasmid DNA isolated from transformants was verified by restriction analysis and the resulting construct was called pGBP02COU-1.

20 pGBP02COU-1 was transformed to *E. coli* JM101 and plasmid DNA isolated from the obtained transformants was verified by restriction analysis. The plasmid DNA was subsequently transformed to *P. freudenreichii* ATCC6207 as described before. Transformants were selected on erythromycin containing agar plates as described before. The plasmid DNA isolated from the *P. freudenreichii*
25 transformants was verified by restriction analysis.

Shake flask fermentations of pGBP02COU-1 transformants using M1 culture medium and the determination of vitamin B12, adenosyl cobinamide and adenosyl cobinamide phosphate levels were performed as described before.

30 Ratios of the amount of vitaminB12, adenosyl cobinamide and adenosyl cobinamide phosphate produced per ml culture OD-end were calculated as described before. The ratio obtained for adenosyl cobinamide was 96% lower than that

obtained for pBRESP36B2p16SH transformants and the ratio obtained for adenosyl cobinamide phosphate was 700% higher than that obtained for pBRESP36B2p16SH transformants. Thus there was a 7-fold increase in the intermediate cobinamide -P when compared with the empty plasmid.

EXAMPLE 8

Construction of an expression vector for the *Propionibacterium freudenreichii* genes encoding protein B and C (cob U and S) and its use for the production of vitamin B12 and its precursors in *Propionibacterium freudenreichii*

In order to over-express simultaneously the genes encoding enzymes B (SEQ ID No.3) and C (SEQ ID No.5) in *Propionibacterium* the genes were placed behind the 16S rRNA promoter and in front of the hemL terminator present in pGBP01HEL-1 as described in the following steps. The two genes are located next to each other on the *P. freudenreichii* genome and are part from the same operon. A DNA fragment containing both genes was amplified by PCR from chromosomal DNA isolated from *P. freudenreichii* ATCC6207 using the following primers: 5'-

CTgATATCAATTggAggACATCAgATgACCCgCATCgTC-3' and 5'-

CTgAATTCCggCggCTCaggCgAACAATg-3'. The first primer contains an EcoRV and a MfeI restriction site and the second primer contains an EcoRI restriction site.

The PCR fragment obtained was ligated into the vector pCR-Blunt II-TOPO (Invitrogen). After transformation of *E. coli* kanamycin resistant colonies were obtained. The DNA sequence of the cloned fragment was verified through sequence analysis. The resulting construct was named pGBPPOB-1.

pGBPPOB-1 was subsequently digested with EcoRV and EcoRI and the fragment containing the genes coding for B and C was isolated and ligated into the large vector fragment isolated from pGBP01HEL-1 which was first digested with ClaI, than blunted with Klenow enzyme, and finally digested with MfeI. After transformation of *E. coli* ampicillin resistant colonies were obtained. Plasmid DNA isolated from transformants was verified by restriction analysis and the resulting

construct was called pGBP02POB-1.

pGBP02POB-1 was transformed to *E. coli* JM101 and plasmid DNA isolated from the obtained transformants was verified by restriction analysis. The plasmid DNA was subsequently transformed to *P. freudenreichii* ATCC6207 as described before. Transformants were selected on erythromycin containing agar plates as described before. The plasmid DNA isolated from the *P. freudenreichii* transformants was verified by restriction analysis.

Shake flask fermentations of pGBP02POB-1 transformants using M1 culture medium and the determination of vitamin B12, adenosyl cobinamide and adenosyl cobinamide phosphate levels were performed as described before.

Ratios of the amount of vitamin B12, adenosyl cobinamide and adenosyl cobinamide phosphate produced per ml culture OD-end were calculated as described before. The ratio obtained for vitB12 was 19% higher than that obtained for pBRESP36B2p16SH transformants. The ratio obtained for adenosyl cobinamide was 96% lower than that obtained for pBRESP36B2p16SH transformants and no adenosyl cobinamide phosphate could be detected in the pGBP02POB-1 transformants.

EXAMPLE 9

Construction of an expression vector for the *Propionibacterium freudenreichii* genes encoding protein B, C and cobA (hence cob A and U and S) and its use for the production of vitamin B12 and its precursors in *Propionibacterium freudenreichii*

In order to over-express simultaneously the genes encoding enzymes B (SEQ ID No.3), C (SEQ ID No.5) and the cobA gene in *Propionibacterium* the three genes were placed behind the 16S rRNA promoter and in front of the hemL terminator as described in the following steps. pGBP02POB-1 was digested with MfeI and BglII and the 1.6 kb fragment was isolated. This fragment was ligated to the 9.7 kb vector fragment of pGBP02COB-1 digested with the same enzymes. After transformation of *E. coli* kanamycin resistant colonies were obtained. The DNA sequence of the cloned

fragment was verified through sequence analysis. The resulting construct was named pGBPPOE-1.

pGBP02POE-1 was transformed to *E. coli* JM101 and plasmid DNA isolated from the obtained transformants was verified by restriction analysis. The plasmid DNA was subsequently transformed to *P. freudenreichii* ATCC6207 as described before. Transformants were selected on erythromycin containing agar plates as described before. The plasmid DNA isolated from the *P. freudenreichii* transformants was verified by restriction analysis.

Shake flask fermentations of pGBP02POE-1 transformants using M1 culture medium and the determination of vitamin B₁₂, adenosyl cobinamide and adenosyl cobinamide phosphate levels were performed as described before.

Ratios of the amount of vitB₁₂, adenosyl cobinamide and adenosyl cobinamide phosphate produced per ml culture OD-end were calculated as described before. The ratio obtained for vitB₁₂ was 45% higher than that obtained for pBRESP36B2p16SH transformants. The ratio obtained for adenosyl cobinamide was 65% lower than that obtained for pBRESP36B2p16SH transformants and no adenosyl cobinamide phosphate could be detected in the pGBP02POE-1 transformants.

SUMMARY OF EXAMPLES

The table below displays the experimental data (the empty plasmid values are set to 100% for reference) for the transformed plasmids relative to the empty plasmid control strain)

Example	Gene	Cobinamide (amount/ml/OD)	Combinamide P (amount/ml/OD)	Vitamin B ₁₂ (amount/ml/OD)
5	Empty plasmid	100	100	100

6	cobA	227 (+127%)	300 (+200%)	94 (-6%)
7	cobU	4 (-96%)	800 (+700%)	94 (-6%)
8	cobU and S	4 (-96%)	0	119 (+19%)
9	CobA and U and S	35 (-65%)	0	145 (+45%)

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